

Results of a palynological study of the contents of small glass bottles of the Late Antiquity from the Kanchaani Cemetery (Southeastern Georgia)

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PL ISSN 0081-3834, e-ISSN: 2719-647X

DOI: <https://doi.org/10.23858/SA/75.2023.2.3150>

<https://rcin.org.pl/dlibra/publication/277430>

Jak cytować:

Kvavadze, E., Martkoplshvili, I., Kakhiani, K., & Rezesidze, N. (2023). Results of a palynological study of the contents of small glass bottles of the Late Antiquity from the Kanchaani Cemetery (Southeastern Georgia). Sprawozdania Archeologiczne, 75(2), 285–309. <https://doi.org/10.23858/SA/75.2023.2.3150>

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RESULTS OF A PALYNOLOGICAL STUDY OF THE CONTENTS OF SMALL GLASS BOTTLES FROM LATE ANTIQUITY FROM THE KANCHAANI CEMETERY (SOUTHEASTERN GEORGIA)

ABSTRACT

Kvavadze E., Martkoplshvili I., Kakhiani K. and Rezesidze N. 2023. Results of a palynological study of the contents of small glass bottles from Late Antiquity from the Kanchaani Cemetery (Southeastern Georgia). *Sprawozdania Archeologiczne* 73/2, 285-309.

Eight small glass bottles (vials) have been obtained from five graves of the Kanchaani cemetery, dating to the 1st-3rd centuries AD. The objects came to the laboratory almost intact, and their contents have also survived. Analysis of the plant pollen and the study of non-pollen palynomorphs (NPP) of these contents showed that there was a set of various medicinal plants in seven bottles, and one bottle contained an infusion made from insects, which also had medicinal properties. The pollen of 23 medicinal plants has been determined to genus and species levels in the contents of the bottles. The paper describes in detail the characteristics of all found medicinal plants and their use in folk medicine. It turns out, that the ethnopharmacology of the Late Antiquity Period in the region under consideration was rather well developed.

Keywords: Late Antiquity Period cemetery, medicinal plants, small glass bottles, Georgia

Received: 13.01.2023; Revised: 07.01.2023; Accepted: 18.01.2023

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1. INTRODUCTION

Palynological examination of the content of the vessels from the Late Antiquity Period of Georgia have never been conducted before, although such analysis has been done of analogous material from the Early Antiquity Period. The evidence studied came from two graves (Nos 22 and 24) of the Vani site located in Western Georgia (Chichinadze *et al.* 2012, 2017, 2019), where organic remains of amphorae and oenochoe were examined. These materials were dated to the 5th-4th centuries BC. Grave 339 of the Pichvnari cemetery, in which the contents of a tumbler were analyzed also dates to the 5th century (Kvavadze and Davadze 2014). From eastern Georgian sites, only the vessel found in Grave 5 of the Khovle necropolis, dated to the 3rd-2nd centuries BC, was studied (Kvavadze and Shatberashvili 2010). It should also be noted, that the vessels from all above mentioned sites are made of clay, with the exception of the oenochoe. The contents of glass vessels of either the Early Antiquity Period or the later period have hardly been studied palynologically either in Georgia or abroad. To this day, it has not been completely clarified what the purpose of this ritual of deposition of these vessels was, namely, what these rather beautiful vials that accompanied the dead into the graves had contained. Most scholars call them scent bottles, which were supposed to have contained the fragrant perfume oil (Lucas and Harris 1962; Colombini *et al.* 2009), but some studies claim that they contained plant infusion or oil for medical purposes (Bardinet 1995; Mortensen 2014).

The Kanchani cemetery is located on the left bank of the river Mashavera on the territory of the village Boslebi, Dmanisi municipality, Kvemo Kartli region, South Georgia. The cemetery consists of two parts - the first (GPS: X 442005.992; Y4576010.687; 980 M.) and



Fig. 1. Map of Georgia and location of the Kanchaani cemetery



Fig. 2. Bottles and vials from the burials of the Kanchaani cemetery: 1 – Grave 2, vial No. 76; 2-5 – Grave 3, bottles No. 84-87; 6 – Grave 4, bottle No. 98; 7 – Grave 12, vial No. 199; 8 – Grave 3, pot No. 82; 9 – Grave 3, pot No. 83

the second (GPS: X 442462.927; Y 4576054.721; 980 M.), (Fig. 1). During the excavation at the cemetery, twenty-five graves were investigated, eight in the first part, seventeen in the second part. All of them are inhumation graves, skeletons were buried in flexed position, on the left or right side, mostly facing west. Among the grave goods were ceramic and glass vessels of various sizes, iron, bronze and gold items, silver coins, beads, *etc.* Based on these material the cemetery in general is dated to the 1st-3rd centuries AD.

Among the grave goods should be distinguished small size glass bottles (Fig. 2). They are almost intact, and their contents have also survived. Six bottles were found in the first part, and two in the second part. They are made of transparent glass. The shape, colour, size as well as the technique of production of each of these bottle varies. One of them was mould-blown, the others were made by the free blowing method. The moulded vessel is light brown and thin-walled, with a high neck, a flat open mouth, a spherical, slightly elongated body, a ring-based bottom, and two ears. Embossed rays extend across the sides of the body. In the free blown vessels, several types are distinguished in shape: jug-like, amphora-like and flask-like (Fig. 2). Such glass vessels were quite often found in the territory of Georgia in Early Antiquity Period graves (Saginashvili 1970; Maisurashvili *et al.* 2018). It should be noted that especially many glass bottles were found at the Urbnisi cemetery. Interestingly here bottles in the burials were placed around the head and in the chest area of the skeletons (Saginashvili 1970).

2. MATERIAL AND METHOD

18 samples of the contents were extracted and palynologically examined from eight small glass bottles and two ceramic pots obtained in the graves of the Kanchaani cemetery. Two samples were taken from each bottle – one from the neck and the other from the bottom. Four small glass vials (Nos 84-87) and two pots (Nos 82 and 83) were found only in Grave 3. In the other four graves, single glass bottles were documented: Bottle No. 76 – Grave 2; Bottle No. 98 – Grave 4; Bottle No. 199 – Grave 12; Bottle No. 203 – Grave 14 (Figs 2-4). Because of the small number of soil samples collected from the neck of the bottles, palynological material almost was not detected in this part of the bottle. The exceptions are the vials No. 84 and No. 98, found in the third and fourth graves (Table 1). In general, the contents are dark organic remains with friable earth. The only exception is the sample from vial No. 199 (Grave 12), the contents of which is of whitish colour.

For the comparative analysis of the palynological spectra, samples were collected from two ceramic vessels within Grave 3. These vessels were selected because they were found next to the glass bottles, together forming a single group for examination.

Material extracted from the contents of vials was processed in a palynological laboratory using the standard chemical method adopted today (Erdtman 1969; Moore *et al.* 1991). At the first stage, the contents were boiled a long time in 10% potassium alkali. At the second stage, the material was centrifuged in a heavy liquid and, finally, acetolysis, that is, colouring, of the material was performed. After washing, the material was placed in glycerol. Palynological study was carried out by means of a new generation light microscope Olympus BX-43. Palynomorphs were photographed using the same microscope.

The results of the palynological research and the analysis of non-pollen palynomorphs are shown on the diagrams (Fig. 3 and 4). The spectra differ from each other by the complexes of plant pollen. Let us discuss peculiarities of each of them.

Kanchaani, NPP

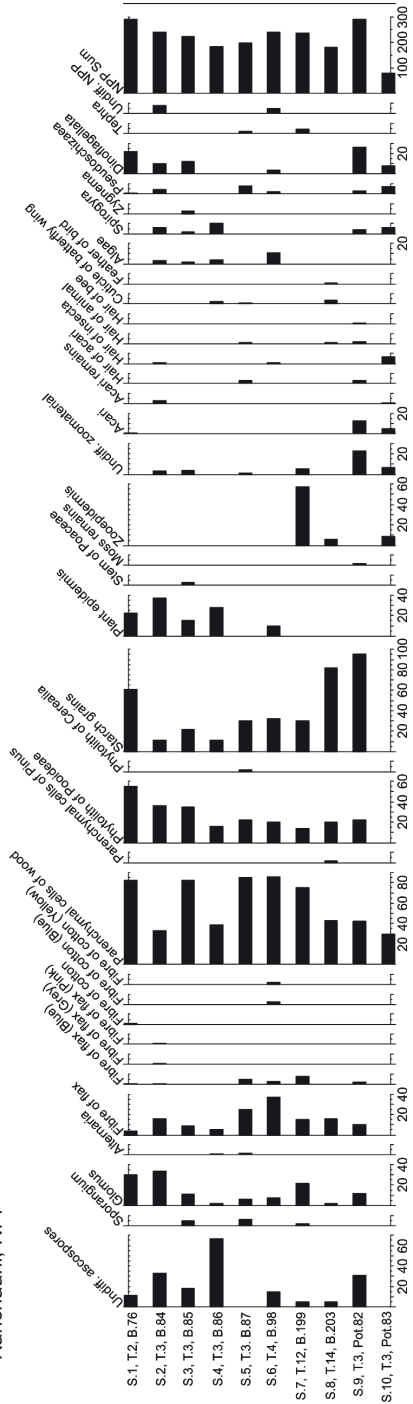


Fig. 4. Non pollen palynomorphs diagram of organic remains of bottles and vials from Kanchaani cemetery.
Explanation: S-sample T – tomb, B – bottle

Table 1. List of samples taken from Kanchaani Cemetery with archaeological and palynological description

| Sample No. | Sample No. on Diagram | Grave No. | Bottle No. | Archaeological context | Palynological context |
|------------|-----------------------|-----------|-------------------------|--|--|
| 1 | 1 | Grave 2 | Neck of Bottle No. 76 | Inhumation burial, crouched on the left side, its head facing north. The grave contained a glass bottle in front of the chest of the skeleton with other goods. | Empty |
| 2 | | | Bottom of bottle No. 76 | | Pollen of 6 plants taxon, 11 types of NPP type |
| 3 | 2 | Grave 3 | Neck of bottle No. 84 | Inhumation burial, crouched on the left side, its head facing south-west. The grave contained four glass bottles in front of the chest of the skeleton with other goods. | Pollen of 2 plants taxon, 4 NPP type |
| 4 | | | Bottom of bottle No. 84 | | Pollen of 6 plants taxon, 14 types of NPP |
| 5 | 3 | Grave 3 | Neck of bottle No. 85 | | Empty |
| 6 | | | Bottom of bottle No. 85 | | Pollen of 9 plants taxon, 14 types of NPP |
| 7 | 4 | Grave 3 | Neck of bottle No. 86 | | Empty |
| 8 | | | Bottom of bottle No. 86 | | Pollen of 6 plants taxon, 11 types of NPP |
| 9 | 5 | Grave 3 | Neck of bottle No. 87 | | Empty |
| 10 | | | Bottom of bottle No. 87 | | Pollen of 11 plants taxon, 14 types of NPP |
| 11 | 9 | Grave 3 | Pot No. 82 | | Pollen of 15 plants taxon, 15 types of NPP |
| 12 | 10 | Grave 3 | Pot No. 83 | | Pollen of 8 plants taxon, 9 types of NPP |
| 13 | 6 | Grave 4 | Neck of bottle No. 98 | Inhumation burial, the grave was damaged, and the position of the skeleton is unclear. The glass bottle was found in the southern part of the grave | Pollen of 2 plants taxon, 3 types of NPP |
| 14 | | | Bottom of bottle No. 98 | | Pollen of 9 plants taxon, 13 types of NPP |

| Sample No. | Sample No. on Diagram | Grave No. | Bottle No. | Archaeological context | Palynological context |
|------------|-----------------------|-----------|--------------------------|---|---|
| 15 | 7 | Grave 12 | Neck of bottle No. 199 | Inhumation burial, crouched on the right side, its head facing south. The grave contained a glass bottle in front of the stomach of the skeleton with other goods | Empty |
| 16 | | | Bottom of bottle 199 | | Pollen of 2 plants taxon, 10 types of NPP |
| 17 | 8 | Grave 14 | Neck of bottle No 203 | The Inhumation burial, Only a fragment of a young person's cranium and a glass bottle were found in the grave. | Empty |
| 18 | | | Bottom of bottle No. 203 | | Pollen of 5 plants taxon, 11 types of NPP |

3. RESULTS

Sample 1, Grave 2, bottle No. 76.

Of woody plants: pollen grains of lime tree (*Tilia*), and pine (*Pinus*) were found in the sample of the contents of this vial. Of herbaceous plants: wormwood (*Artemisia annua*), thistle (*Carduus*), goosefoot (*Chenopodium album*) and forest ferns (*Polypodiaceae*) were detected. Among non-pollen palynomorphs, parenchymal cells of wood predominate (Fig. 4). There were a lot of starch and phytoliths of herbaceous plants. Plant epidermis and fungal spores were well represented as well. There were also spores of fungus *Glomus* in considerable numbers. This fungus grows only on friable soil, and it turned up in this vial from earth, which must have fallen into the bottle after decaying of the cork. There were remains of freshwater algae, ticks and tissue fibres among non-pollen palynomorphs in the examined sample. There were a small quantity of flax fibres, among them also were ones that were light blue in colour. Additionally, cotton fibres were discovered, but they were fewer in number when compared to flax.

Sample 2, Grave 3, bottle No. 84

Four small vials (Nos 84-87) and two pots (Pot Nos 82 and 83) were found here (Fig. 5). From woody plants: pollen of pine (*Pinus*) and currant (*Ribes*) were found in the contents of the bottle. Herbaceous plants were represented by only one species of hawk's-beard (*Crepis aurea*- type). More than 20 pollen grains and 7 pollen clumps of hawk's-beard

were counted, which were found only on the pistil (Fig. 6). Non-pollen palynomorphs were more diverse. Plant epidermis, parenchymal cells of wood and phytoliths of herbaceous plants prevail. Fungal spores, hairs of insects, starch and linen fibres were also found. Remains of freshwater algae were rather well represented.

Pollen grains and clumps of hawk's-beard were also found in the sample taken from the neck of the same bottle. As was already noted, these should be the remains of a hawk's-beard flower. Pollen of common cocklebur was also found in the sample taken from the neck of the bottle. The composition of the non-pollen palynomorphs was nearly the same as in the sample taken from the bottom of the vial. Plant epidermis, phytoliths and wood cells predominate. But fungal spores and linen fibres were more frequent in the sample taken from the neck. Linen fibres of pink, light blue and grey colours were found here.

Sample 3, Grave 3, bottle No. 85

Among woody plants, only pollen of juniper (*Juniperus*) was found in the palynological spectrum of the sample under discussion. In the group of herbaceous plants, the number of pollen grains of plantain (*Plantago major/media*) prevails (Fig. 3). Pollen of cereals, including barley (*Hordeum* type.), was identified. There were also Poaceae. Of sporous plants, spores of adder's-tongue fern (*Opioglossum vulgatum*) and mosses (*Sphagnum*) were found (Fig. 7). Burnt parenchymal cells of wood were abundantly present among the non-pollen palynomorphs. There were a lot of phytoliths of cereals and their starch. Plant epidermis, remains of stalks of cereals, zoo-epidermis and fungal spores were also observed. The following freshwater algae were detected: *Spirogyra*, *Dinoflagellata*, and *Zygnema*. Other algae were also found, the genera of which were not determined at this stage of the study.



Fig. 5. Grave 3, location of bottles Nos 84-87 and pots Nos 82 and 83

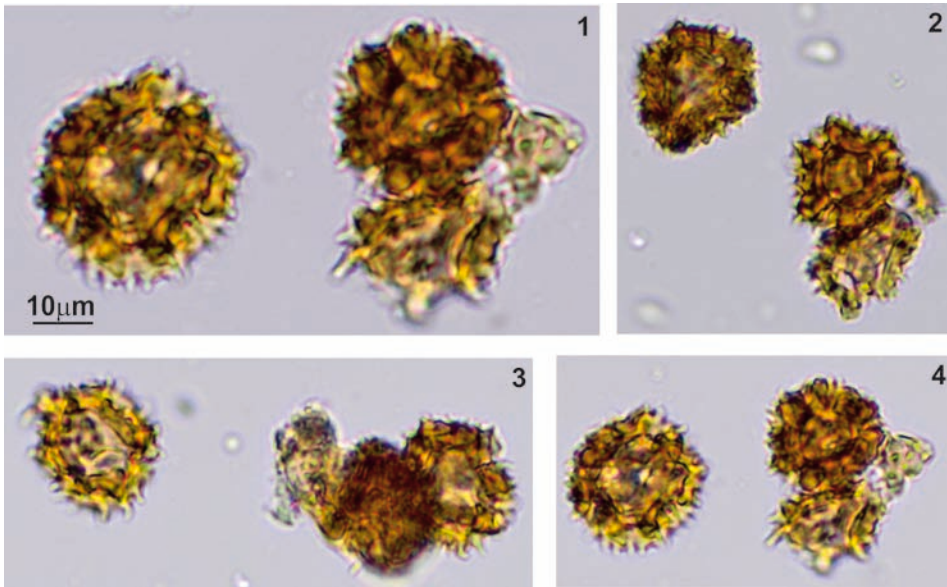


Fig. 6. Grave 3, bottle No. 84: 1-4 – *Crepis* and clumps of *Crepis* pollen

Sample 4, Grave 3, bottle No. 86.

This is a small bottle with a very narrow neck (Fig. 2). Of woody plants: pollen grains of pine (*Pinus*), spruce (*Picea*), alder (*Alnus incana*-type) and vine (*Vitis vinifera*) were found. It should also be said that plant pollen grains were well preserved (Fig. 8). From the herbaceous plants: there were only wormwood (*Artemisia*, *Artemisia annua*-type) and grasses (Poaceae). Fungal spores and tracheal cells of wood predominate among non-pollen palynomorphs. There was a lot of plant epidermis, phytoliths and starch. Remains of freshwater algae (*Spirogyra*) were also well represented. Fibres of flax textile were observed as well.

Sample 5, Grave 3, bottle No. 87

Of woody plants: pollen grains of juniper (*Juniperus*), pine (*Pinus*), birch (*Betula*) and oak (*Quercus*) were found in the contents of this vial, and of herbaceous plants: those of apium (*Apium*), wormwood (*Artemisia*), hairy bittercress (*Cardamine hirsuta*), thistle (*Carduus*), common mouse-ear chickweed (*Cerastium*), stinging nettle (*Urtica*), and cannabis (*Cannabis sativa*). In non-pollen palynomorphs, parenchymal cells of burnt wood prevail here as well. There were a lot of grains of cereal starch and their phytoliths. Wheat phytoliths were also observed. Fungal spores, remains of ticks and insects were represented in a small quantity. Hairs of animal was also found, as well as algae *Pseudoschizaea*. There was a rather large amount of linen textile fibres. Among them fibres dyed blue and light blue were detected.

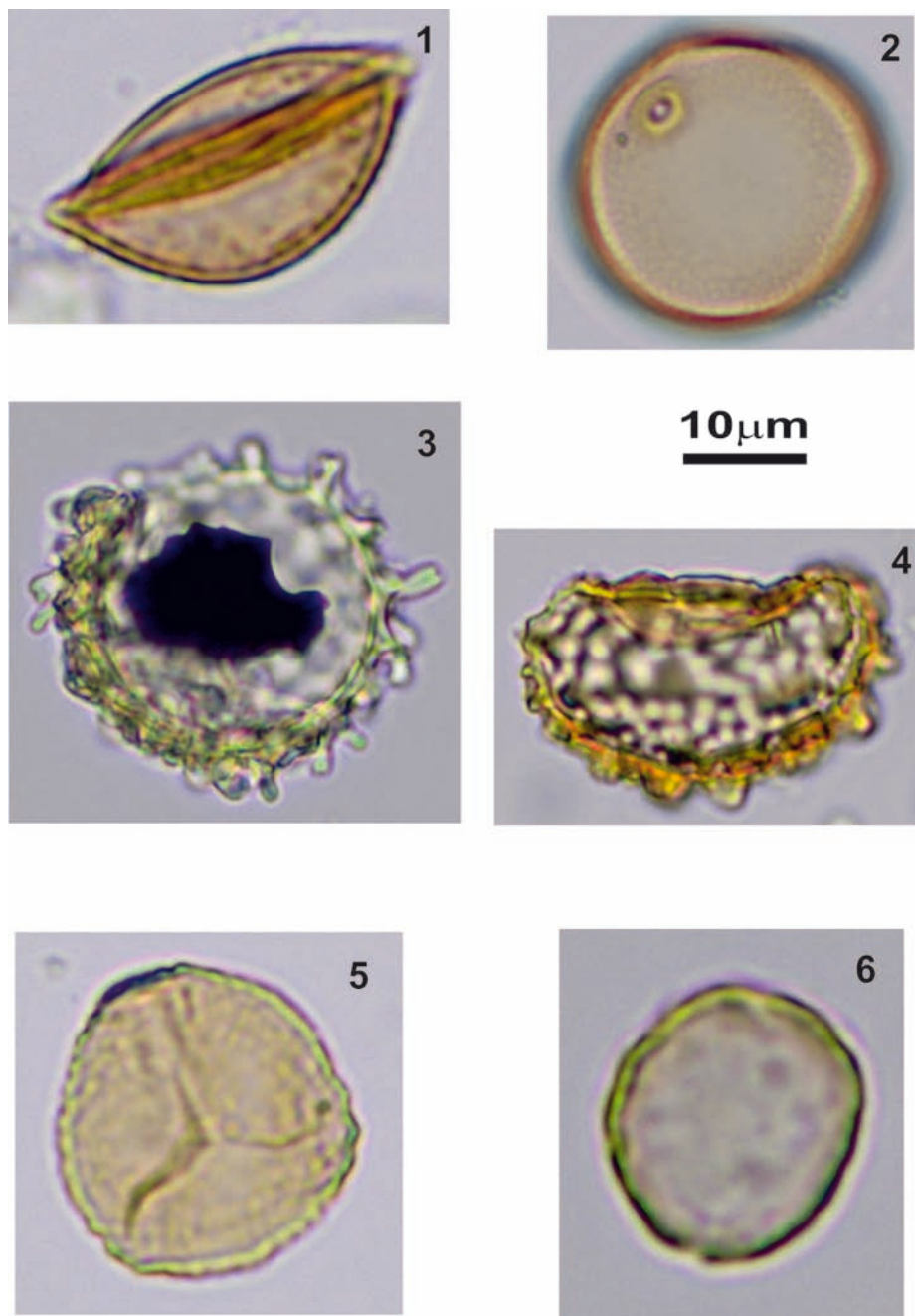


Fig. 7. Grave 3, bottle No. 85: 1 – *Juniperus*; 2 – *Hordeum* type; 3,4 – *Ophioglossum vulgatum*; 5 – *Sphagnum*; 6 – *Cannabis sativa*

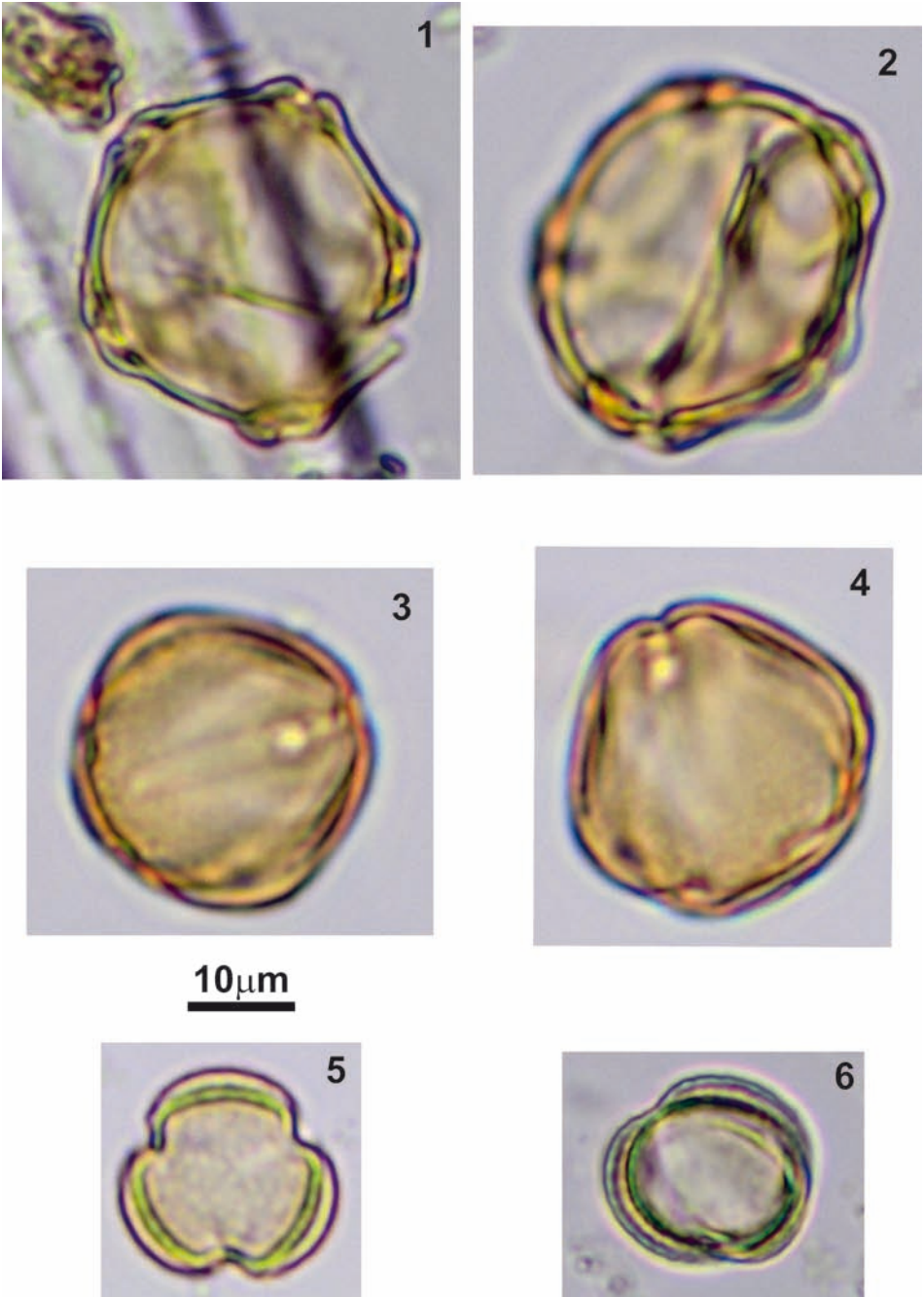


Fig. 8. Grave 3, bottle No. 86: 1, 2 – *Alnus*; 3, 4 – *Vitis vinifera*; 5, 6 – *Artemisia*

Sample 6, Grave 4, bottle No. 98

In the contents of the bottle only vine pollen grains (*Vitis vinifera*) were found in the group of woody plants and bushes. There were more herbaceous plants here, among them pollen grain clumps of hawk's-beard (*Crepis aureotype*), common cocklebur (*Xanthium*), plants of the pink family (Caryophyllaceae), grasses (*Poaceae*) and other undetermined tricollporate pollen. Parenchymal cells of wood and vine starch prevail among the non-pollen palynomorphs. Among textile fibres, linen predominated, which was well represented. Linen and cotton fibres, were dyed light blue. Some yellow cotton fibres were also found. Besides, fungal spores, phytoliths of cereals and freshwater algae were identified.

Plant pollen was almost absent in the sample taken from the neck of the same bottle, but there were a lot of parenchymal cells of wood and linen fibres. Plant epidermis, fungal spores, starch, phytoliths and remains of freshwater algae were present in a small amount. Therefore, we can suppose that we have the remains of a wooden cork in the neck of the vial. Linen was wrapped around the cork, as there were a lot of wood cells and textile fibres in this sample.

Sample 7, Grave 12, bottle No. 199

The contents were left only in the big and elongated neck of the vial under discussion, as it was found in an almost horizontal position and placed by the abdomen of the deceased. This bottle resembles more a chemical measuring glass. It is of big size and greenish in colour. The palynological spectrum contains only one pollen grain of pine and two pollen grains of representatives of the genus *Cichorium*. Cells of wood and epidermis of insects and their microscopic remains of other types prevail among the non-pollen palynomorphs. About 60 insect remains have been counted, that was not recorded in any other vials. That's why we assume that some kind of infusion made of insects was kept in this bottle. It should also be noted that remains of volcanic ashes were found only in the contents in question. Fungal spores, starch and phytoliths were also found in the contents of the vial neck. Linen fibres and among them those dyed light blue were well represented. There were freshwater algae *Spirogyra* and *Pseudoschizaeae* in small numbers.

Sample 8, Grave 14, bottle No. 203

This is a small vessel with a narrow neck. Its contents were dark and dense (non friable). Of woody plants, pollen grains of pine (*Pinus*), lime tree (*Tilia*) and vine (*Vitis vinifera*) were found in the palynological spectrum. Of herbaceous plants, only pollen grains of goosefoot and spores of forest ferns were detected. Starch, including that of vine, predominates in the non-pollen palynomorphs. There were a lot of parenchymal cells of charred wood and among them cells of pine (*Pinus*) (Fig. 9). Phytoliths were well represented. Small amounts of fungal spores, bird feathers, animal hair and scales of butterfly wings were found. It should also be noted that dark cells of wood were found in the content of the vial, which were microscopic traces of ashes or smoke, as well as light ones that should be the remains of a cork. Only pine cells were of light colour here, so we might assume that the cork was made of pine tree. The cork must have been wrapped in a piece of linen here as well, as there were a lot of linen fibres in the neck (Fig. 3).

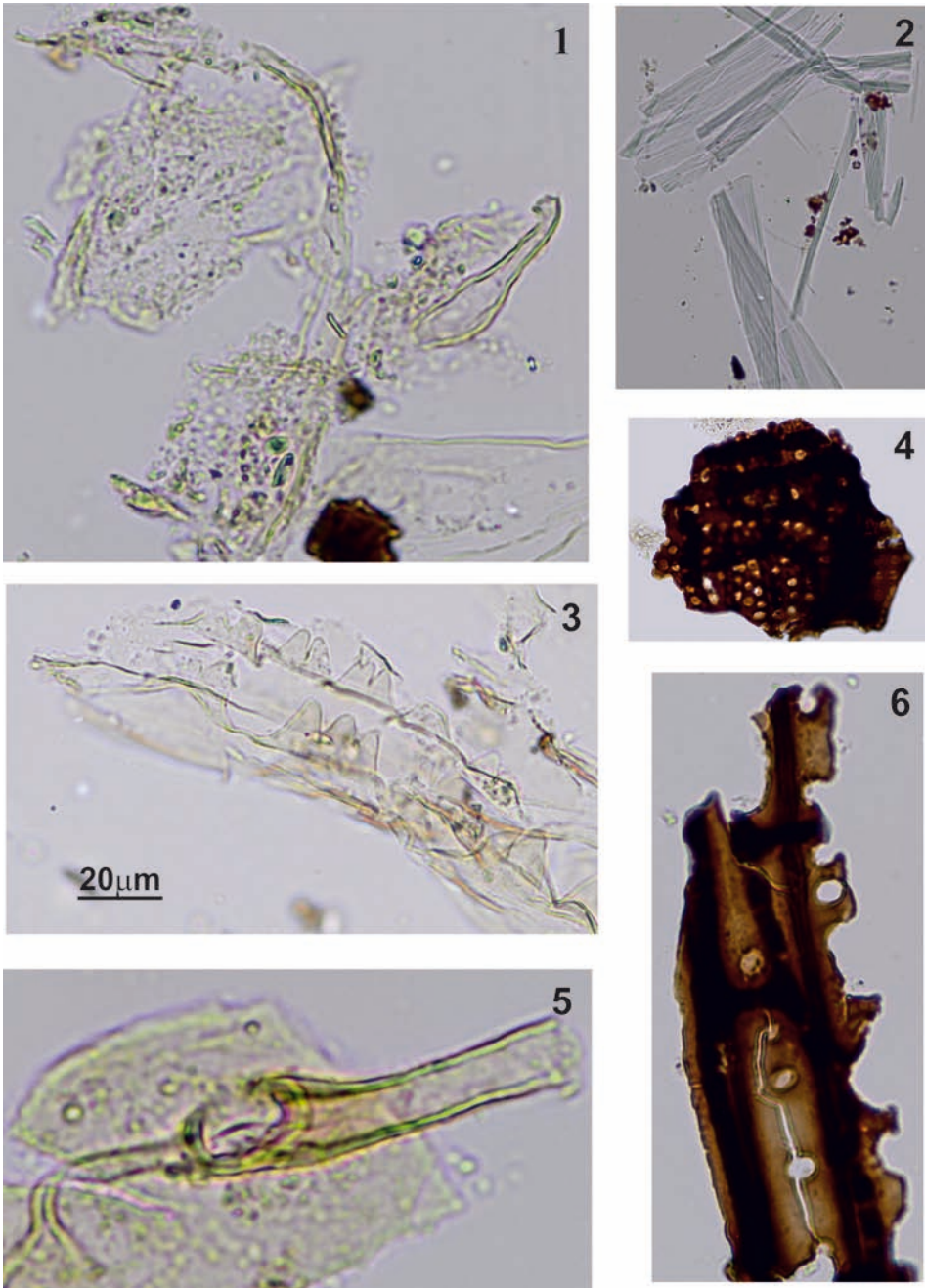


Fig. 9. Grave 12, vial No. 199: 1, 3, 4 – insects epidermis; 2 – flax textile fibres; 4 – tracheal cells of undiff. wood; 5 – Grave 14, vial 203: insect epidermis; 6 – parenchymal cells of *Pinus* (pine) from cork

Sample 9-10, Grave 3, pots Nos 82 and 83

As has been already noted, the contents of two clay vessels from the Grave 3 were also examined. They were both small jugs, approximately, about one litre in volume (Fig. 2). The palynological spectrum of the content of the vessel with a broken handle (Pot No. 82) was rather rich in the composition of pollen grains (Fig. 3). Among woody plants: pollen grains of hornbeam (*Carpinus betulus*) and pine (*Pinus*) prevail. The pollen of oak (*Quercus*), beech (*Fagus orientalis*), walnut (*Juglans regia*) and hazel (*Corylus*) was observed. The quantity of herbaceous plants in the spectrum was less than the above mentioned trees. This group also contains pollen of some cereals. These were wheat, absinth, thistle, Iranian knapweed, goosefoot, and spores of forest ferns. Starch prevails in non-pollen palynomorphs. There were a lot of wood cells, fungal spores, phytoliths. Remains of insects and other invertebrates, as well as algae, were well represented. There was a small amount of linen fibres, moss residues and spores of the fungus *Glomus*, indicating contamination from the soil. Pot No. 83 from Grave 3 is characterized by a completely different palynological spectrum. There was almost no pollen of herbaceous plants. From tree plants, separate pollen grains of pine, oak, hornbeam, walnut, alder, birch, lime tree and hazel were present in a small amount (Fig. 3). From non-pollen palynomorphs, only cells of wood, insect remains and freshwater algae were found (Fig. 4).

4. DISCUSSION

The palynological spectra of the material under discussion differ both in the complexes of plant pollen grain and in the quantity of non-pollen palynomorphs. This picture is well seen on the palynological diagram (Figs 3 and 4). It is also noticeable that the spectra of the big-sized vials with a wide mouth (Nos 76, 84 and 199) are distinguished by a large number of spores of the fungus *Glomus*. This indicates a greater contamination of its contents with grave soil. As has already been mentioned, this microscopic fungus grows only in the loose soil (Van Geel 1998; Geel *et al.* 2003; Geel and Aptroot 2006). The least amount of the spores of this fungus was detected in the small and narrow necked vessel (Nos 86-88 and 203), where the soil of the grave walls could not penetrate so easily into the vessel. The same situation is found with the number of phytoliths. There are more of them in large and wide-mouthed vials than in the ones with narrow necks. Phytoliths consist of silica, so they also are well preserved in the soil (Piperno 2006).

The study and analysis of the plant composition of those palynological spectra that are less contaminated, showed that they consisted completely of plants with medicinal uses. Table 2 provides a detailed breakdown of the diseases for which these plants have been used. It's worth noting that only the members of the pink family (Caryophyllaceae) are odorous within the spectrum, and nearly all of them possess medicinal properties (Chandra and Rawat 2015). Among trees and shrubs, nine medicinal plants are evident. Notably,

pollen grains of pine were discovered in five of the bottles under consideration. In traditional medicine, pine flowers, cones, needles, and at times its resin are used to create remedies for respiratory diseases such as asthma, pneumonia, and tuberculosis. (Khare 2007; Alarcon *et al.* 2015; Bussmann *et al.* 2016). The pollen of juniper (which also belongs to the coniferous plants) was found in two bottles (Nos 85 and 97). Juniper can be used as a remedy for epilepsy (Adams *et al.* 2012; Alarcon *et al.* 2015), as well as an antiseptic (Alarcon *et al.* 2015) in ethnopharmacology. It is also used as an antibacterial agent, a remedy for hepatitis, diabetes, helminthiasis and other diseases (Swanston-Flatt *et al.* 1990; Tilford 1997; Al-Snafi 2018). The medicine is made from almost any part of the juniper, namely from the fruits, cones, needles, wood. Healing oil is also made from it, as its wood contains oil (Al-Snafi 2018). It should also be noted that juniper pollen grains are poorly preserved in the soil, as it has a very thin exine (Kurmann 1994). It can be assumed that both vials in which juniper pollen was found in better condition (Fig. 7), were used to keep juniper oil. Another argument is that it was in these bottles that the largest amount of pollen of medicinal plants was found (Table 1). The oil preserved pollen of other eight plants in bottle No. 85 and that of nine plants in bottle No. 87. The diagram shows that in other bottles, except for No. 89, there are much fewer taxa. Therefore, we can suppose that the bottles would have contained oil, which preserved not only grains of juniper pollen but that of other plants as well. The vials are of a small size, which is another argument in favour of the fact that they would have contained oil, each drop of which had healing properties.

Oil must have been kept in the bottle No. 98, where vine pollen is also presented. Like juniper pollen grains, vine pollen is hardly ever preserved in the soil (Kvavadze *et al.* 2010; McGovern *et al.* 2017). Therefore it should be proposed that oil in the bottle No. 98 was pressed from grapeseeds, which are always followed by pollen grains (Tvalchrelidze and Kvavadze 2016). The number of plant taxa (nine) is also large in the palynological spectrum in question. It should also be noted that all of them are of medicinal plants (Table 2).

The palynological spectrum of the bottle No. 84 is of great interest. It contains a large number of pollen of hawk's-beard (*Crepis*), including numerous clumps of pollen (Fig. 6). This suggests that there would have been an infusion of the flower of this plant. Its pollen, like other Cichorioideae, is preserved well and possibly here we are dealing with traces of an infusion of flowers boiled in water. There are also many algal residues in the spectrum (Fig. 4). Hawk's-beard (*Crepis*) treats hepatitis. It is used as an anti-inflammatory and antioxidant agent (Fleurentin *et al.* 1986; Quattrocchi 2012; Pedreiro *et al.* 2021). Hawk's-beard (*Crepis*) was also found in the bottle No. 98, which would have contained vine oil. It was also identified in the content of the jug No. 82. Several species of hawk's-beard (*Crepis*) still grow in the territory of Georgia, and 38 of its species are widespread throughout the Caucasus (Grosgeim 1949).

As was already noted, in the vessel No. 199 from Grave 12, only remains of insects were found, that means that their infusion or decoction would have been in it (Fig. 9). Insect remains were also found in both clay jugs from the third grave. In the traditional medicine

Table 2. List of medicinal plants of the Late Antiquity Period found in the small glass bottles from Kanchaani cemetery and their pharmacological properties

| Plant | Family | Pharmacological Properties | References |
|---|-----------------|---|--|
| <i>Pinus</i> (pine) | Pinaceae | Asthma, rheumatism, antiseptic, pneumonia, cystitis, painkiller, wounds | Lim 2012b; Kizirarslan and Sevğ 2013; Alarcün <i>et al.</i> 2015 |
| <i>Juniperus</i> (juniper) | Cupressaceae | Epilepsy, gastrointestinal diseases, antiseptic, diarrhea, diuretic, earache | Swanston-Flatt <i>et al.</i> 1990; Tilford 1997; Al-Snafi 2018 |
| <i>Betula</i> (birch) | Betulaceae | Antipyretic, epilepsy, painkiller, cystitis, rheumatism | Adams <i>et al.</i> 2012; Papp <i>et al.</i> 2014; Al-Snafi 2015 |
| <i>Quercus</i> (oak) | Fagaceae | Wounds, hypertension, preventing a cold, diabetes, antimicrobial | Keskin and Alpınar 2002; Mikaili <i>et al.</i> 2012; Taib <i>et al.</i> 2020 |
| <i>Alnus</i> (alder) | Betulaceae | Anthelmintic, diarrhoea, dysentery, gastritis, hemorrhage | Quattrocchi 2012; Menale and Mouio 2014; Dahija <i>et al.</i> 2016 |
| <i>Tilia</i> (lime) | Tiliaceae | Preventing a cold, cough, high temperature, antiseptic, hemorrhage, cystitis | Demiray 2009; Güler <i>et al.</i> 2015; Poljšak and Glavač 2021 |
| <i>Vitis vinifera</i> (common grape vine) | Vitaceae | Epilepsy, anemia, painkiller, antiallergic, preventing a cold, cancer, toothache, antibacterial | Adams <i>et al.</i> 2012; Kanagarla <i>et al.</i> 2013; Hayta <i>et al.</i> 2014 |
| <i>Ribes</i> (currants) | Grossulariaceae | Diabetes disease, kidney stones; analgesic; antiseptic | Duke 2002; Hayta <i>et al.</i> 2014; Cao <i>et al.</i> 2021 |
| <i>Hordeum</i> (barley) | Poaceae | Hepatitis, abdominal pains, cough, diarrhoea, painkiller, toothache | Chevallier 1996; Marwat <i>et al.</i> 2012; Lim 2013 |
| <i>Crepis</i> (hawksbeard) | Asteraceae | Anti-inflammatory, antioxidant | Fleurentin <i>et al.</i> 1986; Quattrocchi 2012; Pedreiro <i>et al.</i> 2021 |
| <i>Artemisia</i> (fragrant wormwood) | Asteraceae | Malaria, fever, antiseptic, rheumatism, anthelmintic, diarrhoea, diabetes, anti-bug | Hayta <i>et al.</i> 2014; Bussmann <i>et al.</i> 2016; Koul <i>et al.</i> 2017 |
| <i>Artemisia annua</i> (wormwood) | Asteraceae | Malaria, antiseptic, fever, anti-bug | Liu <i>et al.</i> 1992; Ferreira 2004; Mueller <i>et al.</i> 2004; de Ridder <i>et al.</i> 2008; |
| <i>Xanthium</i> (cocklebur) | Asteraceae | Antimicrobial, gastrointestinal diseases, antibacterial, preventing a cold, rhinitis, toothache | Eissa <i>et al.</i> 2013; Li and Xing 2016; Khan <i>et al.</i> 2020 |
| <i>Carduus</i> (thistle) | Asteraceae | Gastrointestinal diseases, snakebite, antipyretic, causing vomiting | Dold and Cocks 2000; Zheleva-Dimitrova <i>et al.</i> 2011; Hayta <i>et al.</i> 2014 |
| <i>Chenopodium album</i> (goosefoot) | Chenopodiaceae | Anthelmintic, diuretic, arthritis, rheumatism, scurvy, talc | Meuninck 2013; Bibi <i>et al.</i> 2014; Trivedi and Singh 2018 |
| <i>Urtica</i> (stinging nettle) | Urticaceae | Rheumatism, asthma, hemorrhage, high temperature, mumps, rhinitis, anemia, fungal diseases | Kültür <i>et al.</i> 2007; Dyubeni and Buwa 2012; Kregiel <i>et al.</i> 2018 |
| <i>Cannabis sativa</i> (ordinary hemp) | Cannabaceae | Sensation of dizziness, diarrhoea, fracture, swelling, painkiller, anthelmintic | Kalant 2001; Bibi <i>et al.</i> 2014; Jamila and Mostafa 2014 |
| <i>Apium</i> (celery) | Apiaceae | Antispasmodic, diuretic, rheumatism, gout, painkiller, diarrhoea, dysentery | Leto <i>et al.</i> 2013; Kooti <i>et al.</i> 2014; Alarcün <i>et al.</i> 2015 |

| Plant | Family | Pharmacological Properties | References |
|--|-----------------|--|--|
| <i>Cerastium</i> (chickweed) | Caryophyllaceae | Headache, renal colic, body ache, decoction, cough | Angmo <i>et al.</i> 2012; Chandra and Rawati 2015; |
| <i>Cardamine</i> (toothworts) | Brassicaceae | Dysentery, epilepsy; diuretic, depurative, antispasmodic | Wiar 2006; Hatfield 2004; Quatrocchi 2012 |
| <i>Plantago</i> (great plantain) | Plantaginaceae | Antiseptic, skin diseases, arthritis, diseases of respiratory ways, gastrointestinal diseases, burning, hemorrhage | Kültür <i>et al.</i> 2007; Tetik <i>et al.</i> 2013; Najafian <i>et al.</i> 2018 |
| <i>Ophioglossum vulgatum</i> (adder's tongue) | Ophioglossaceae | Antiseptical, hemorrhage, angina, hematoma, vomiting, wounds, antiviral, treating frostbite, burning | Mannan <i>et al.</i> 2008; Quatrocchi 2012; Lim 2016a; 2016b. |
| <i>Sphagnum</i> (sphagnum moss) | Sphagnaceae | Hot compress, abscess characteristic of diseases of urogenital systems, eye lotion | Souter 1995; Allen and Hatfield 2004; Meuninck 2013 |

of many countries around the world, the treatment of several diseases using insect extracts or decoctions is a well-established practice. For example, in China there are more than hundred such kind of insects (Feng *et al.* 2009). Today there is a special branch of medicine called “entomotherapy”, that is entomological therapy “Insect-derived compounds for therapeutic use have huge promise as they have remedial, analgesic, antibacterial, diuretic, sedative, anticancer and anti-rheumatic properties” (Kaur *et al.* 2023). Insect infusions possess antibacterial, antianalgesic properties, treats diarrhea, rheumatism and other diseases (Yamakawa 1998; Costa-Neto 2002, 2005; Bairagi 2019; Mozhui *et al.* 2021).

The fact that the vessel with insect infusion in Grave 12 was placed near the abdomen of the deceased might suggest that he suffered from diarrhoea, and that is why the vessel full of remedy was placed near the abdomen (Fig. 10).

Palynological examination of the content of eight vials has shown that each each vessel contained an infusion or decoction of various medicinal plants. The pollen of 23 medicinal herbs have been determined to genus and species, which indicates that the population inhabiting the territory of Georgia in the Late Antiquity Period very well knew the properties of numerous plants. At that time, the medicinal properties of insects were also well known, that is why three vessels with medicines made of insects accompanied the dead in the graves.

It is interesting to note that all the bottles had a wooden cork. These corks would have been made of pine wood (Richter *et al.* 2004) and wrapped in a piece of linen for greater safety, or probably corks were only made of textile. This is evidenced by the presence of wood parenchymal cells and large amounts of flax fibres in a spectrum of non-pollen palynomorphs. Among the wooden cells there are light cells that did not burn out in the fire, but rotted naturally. The cells that were burnt in fire are of much darker in colour, than the parenchymal cells of a decayed tree (Kvavadze and Kakhiani 2010). Based on this difference, it is possible to determine the presence of decoction and tincture of medicinal plants. In the case if there is a healing tincture, in the spectrum of non-pollen palynomorphs, light-



Fig. 10. Grave 12, location of vial No. 199

coloured, decomposed wood parenchymal cells predominate, which are the remains of the cork. In the case of the decoction, the spectrum is dominated by the burnt wood parenchymal cells.

5. CONCLUSION

The palynological study and, in particular, the existence of non-pollen palynomorphs have shown that small glass bottles placed in graves were tightly closed from the very beginning with corks that were wrapped with a piece of cloth. This circumstance has preserved all the components of the contents in the bottles. These are pollen of medicinal plants, remains of freshwater algae, which had existed in the water in which plants were boiled. During boiling, sterilization also took place, which subsequently contributed to the preservation of pollen grains and microscopic remains of other organisms. In addition, in four bottles, where the preservation was particularly good, traces of vine seed oil and juniper fruit or wood oil were found. It is well known that oil is an excellent conservation agent (as fungi, microbes or bacteria that destroy the organic remains do not thrive in it).

Palynological analysis has revealed the presence of infusion and decoction of various medicinal plants in eight bottles. The pollen of 23 medicinal plants was detected. This suggests that during the Late Antiquity Period, the population living near the Kanchaani cemetery knew the medicinal properties of several plants. Moreover, during the 1st-3rd centuries AD, the population was also aware of the medical properties of insects, which were placed in the graves of two individuals as part of the funerary rites.

References

- Adams M., Schneider S. V., Kluge M., Kessler M. and Hamburger M. 2012. Epilepsy in the Renaissance: A survey of remedies from 16th and 17th century German herbals. *Journal of Ethnopharmacology* 143, 1-13.
- Alarcón R., Pardo-de-Santayana M., Priestley C., Morales R. and Heinrich M. 2015. Medicinal and local food plants in the south of Alava (Basque Country, Spain). *Journal of Ethnopharmacology* 176, 207-224.
- Al-Snafi A. 2015. The medical importance of *Betula alba* – An overview. *Journal of Pharmaceutical Biology* 5/2, 99-103.
- Al-Snafi A. 2018. Medical importance of *Juniperus communis* – A Review. *Indo American Journal of Pharmaceutical Sciences* 5/3, 1779-1792.
- Allen D. E. and Hatfield G. 2004. *Medicinal plants in folk tradition*. Cambridge, Portland: Timber Press.
- Angmo K., Adhikari B. S. and Rawat G. S. 2012. Changing aspects of traditional healthcare system in Western Ladakh, India. *Journal of Ethnopharmacology* 143/2, 621-630.
- Bairagi S. H. 2019. Insects with Potential Medicinal Significance: A Review. BIOMED, *Journal of Scientific & Technical Research* 16/3, 12024-12027.
- Bardinet T. 1995. *Le Papyrus Médicaux de l'Égypte Pharaonique*. Paris: Fayard.
- Bibi S., Sultana J., Sultana H. and Malik R.N. 2014. Ethnobotanical uses of medicinal plants in the highlands of Soan Valley, Salt Range, Pakistan. *Journal of Ethnopharmacology* 155, 352-361.
- Busmann R. W., Paniagua Zambrana N. Y., Sikharulidze S., Kikvidze Z., Kikodze D., Tchelidze D., Batsatsashvili K. and Robbie H. E. 2016. Medicinal and Food Plants of Svaneti and Lechkhumi, Sakartvelo (Republic of Georgia), Caucasus. *Medicinal & Aromatic Plants* 5, 266. doi: 10.4172/2167-0412.1000266
- Cao L., Park Y., Lee S. and Kim D. O. 2021. Extraction, Identification and Health Benefits of Anthocyanins in Blackcurrants (*Ribes nigrum* L.). *Applied Sciences* 11/4, 1863. <https://doi.org/10.3390/app11041863>
- Chandra S. and Rawat D. S. 2015. Medicinal plants of the family Caryophyllaceae: a review of ethno-medicinal uses and pharmacological properties. *Integrative Medicine Research* 4, 123-131.
- Chevallier A. 1996. *The encyclopedia of medicinal plants*. London: Dorling Kindersley.
- Chichinadze M., Kvavadze E., Kacharava D., Akhvlediani D. and Kvirkvelia G. 2012. Palynological characteristics of organic remains of Tomb no. 22 from the site of the ancient settlement Vani. *Proceeding of Natural and Prehistoric Section, Georgian National Museum* 4, 95-108 (in Georgian).
- Chichinadze M., Kvavadze E. and Martkoplshvili I. 2017. Environmental conditions at the Vani Site of the Classical period according to palynological data. *Bulletin of The Georgian National Academy of Sciences* 11/4, 112-118.
- Chichinadze M., Kvavadze E., Martkoplshvili I. and Kacharava D. 2019. Palynological evidence for the use of honey in funerary rites during the Classical Period at the Vani. *Quaternary International* 507, 24-33.

- Colombini M. P., Giachi G. and Ribechini M. I. E 2009. An Etruscan ointment from Chiusi (Tuscany, Italy): its chemical characterization. *Journal of Archaeological Science* 36, 1488-1495.
- Costa-Neto E. M. 2002. The Use of Insects in Folk Medicine in the State of Bahia, Northeastern Brazil, with Notes on Insects Reported Elsewhere in Brazilian Folk Medicine. *Human Ecology* 30, 245-263.
- Costa-Neto E. M. 2005. Entemotherapy, or the medicinal use of insects. *Journal of Ethnobiology* 25/1, 93-114.
- Dahija S., Haverić S., Čakar J. and Parić A. 2016. Antimicrobial and cytotoxic activity of *Alnus glutinosa* (L.) Gaertn. *A. incana* (L.) Moench, and *A. viridis* (Chaix) DC. extracts. *Journal of Health Sciences* 6/2, 100-104.
- De Ridder S., van der Kooy F. and Robert Verpoorte R. 2008. *Artemisia annua* as a self-reliant treatment for malaria in developing countries. *Journal of Ethnopharmacology* 120, 302-314.
- Demiray S., Pintado M. E. and Castro P. M. L. 2009. Evaluation of phenolic profiles and antioxidant activities of Turkish medicinal plants: *Tilia argentea*, *Crataegi folium* leaves and *Polygonum bistorta* roots. *World Academy of Science, Engineering and Technology International Journal of Medical, Health, Biomedical, Bioengineering and Pharmaceutical Engineering* 3/6, 74-79.
- Dold A. P. and Cocks M. L. 2000. The medicinal use of some weeds, problem and alien plants in the Grahamstown and peddie districts of the Eastern Cape, South Africa. *South African Journal of Science* 96, 467-473.
- Dyubeni L. and Buwa L. V. 2012. An ethnobotanical study of plants used for the treatment of ear, nose and throat (ENT) infections in Nkonkobe Municipality. South Africa. *Journal of Medicinal Plants Research* 6(14), 2721-2726.
- Duke J. A., Bogenschutz-Godvin M. J., du Cellier L. and Duke P-A. K. 2002. *Handbook of medicinal plants*, second edition. Boca Raton London New York Washington, D.C. Boca Raton London New York Washington, D.C: CRC Press.
- Eissa T. A. F., Palomino O. M., Carretero M. E. and Gómez-Serranillos M. P. 2013. Ethnopharmacological study of medicinal plants used in the treatment of CNS disorders in Sinai Peninsula, Egypt. *Journal of Ethnopharmacology* 151, 317-332.
- Erdtman G. 1969. *Handbook of Palynology*. Copenhagen: Munksgaard.
- Feng Y., Zhao M., Zhao HE., Chen Z. and Sun L. 2009. Research and utilization of medicinal insects in China. *Entomological Research* 39/5, 313-316.
- Ferreira J. F. S. 2004. *Artemisia annua* L.: The hope against malaria and cancer. In *Proceedings of the Meeting on Medicinal and Aromatic Plants: Production, Business & Applications*, January 15-7, Beckley. WV: Mountain State University.
- Fleurentin J., Mynnti C. and Pelt J. M. 1986. Traditional medicine and traditional healers in North Yemen. *Curare* 5, 133-144.
- Grossgeim G. G. 1949. *The identifier of the flora of the Caucasus*. Moscow: Sovetskaya Nauka.
- Güler B., Manav E. and Uđurlu E. 2015. Medicinal plants used by traditional healers in Bozüyük (Bilecik-Turkey). *Journal of Ethnopharmacology* 173, 39-47.
- Hayta S., Polat R. and Selvi S. 2014 Traditional uses of medicinal plants in Elazýd (Turkey). *Journal of Ethnopharmacology* 154, 613-623.

- Hatfield G. 2004. *Encyclopedia of folk medicine: old world and new world traditions*. California: ABC-CLIO.
- Jamila F. and Mostafa E. 2014. Ethnobotanical survey of medicinal plants used by people in Oriental Marocco to manage various ailments. *Journal of Ethnopharmacology* 154, 76-87.
- Kalant H. 2001. Medicinal Use of Cannabis: History and Current Status. *Pain Research and Management* 6/2, 80-91.
- Kanagarla N., Kuppast I. J., Veerashekar T. and Reddy C. L. 2013. A review on benefits and uses of *Vitis vinifera* (Grape). *Research and reviews in biosciences* 7/5, 175-180.
- Kaur G., Sarao P. S. and Chhabra N. 2023. Therapeutic use of insect and insect products. *Indian Journal of Entomology* Online published Ref. No. e23964, 1-10.
- Keskin M. and Alpýnar K. 2002. An ethnobotanical study on Kýtlak (Yayladađý-Hatay). *The Herb Journal of Systematic Botany* 9, 91-100.
- Khan Y., Shah S. and Ullah Sh. 2020. Ethnomedicinal, pharmacological and phytochemical evaluation of *Xanthium strumarium* L. *International Journal of Scientific & Engineering Research* 11/7, 587-591.
- Khare C.P. 2007. *Indian medicinal plants. An illustrated dictionary*. Verlag Berlin/Heidelberg/ Kultur: Springer.
- Kýzýlarslan Ç. and Sevg E. 2013. Ethnobotanical uses of genus *Pinus* L. (Pinaceae) in Turkey. *Indian Journal of Traditional Knowledge* 12/2, 209-220.
- Kooti W., Aliakbari S., Asadi-Samani M., Ghadery H. and Ashtary-Larky D. 2014. A review on medicinal plant of *Apium graveolens*. *Advanced Herbal Medicine* 1/1, 48-59.
- Koul B., Taak P., Kumar A., Khatri T. T. and Sanyal I. 2018. The *Artemisia* Genus: A Review on Traditional Uses, Phytochemical Constituents, Pharmacological Properties and Germplasm Conservation. *Journal of Glycomics & Lipidomics* 7, 1-7.
- Kregiel D., Pawlikowska E. and Antolak H. 2018. *Urtica* spp.: Ordinary Plants with Extraordinary Properties. *Molecules* 23, 1664. <https://doi.org/10.3390/molecules23071664>
- Kültür S. 2007. Medicinal plants used in Kýrklareli Province (Turkey). *Journal of Ethnopharmacology* 111, 341-364.
- Kurmann M. H. 1994. Pollen morphology and ultrastructure in the Cupressaceae. *Acta Botanica Gallica* 141/2, 141-147.
- Kvavadze E. and Kakhiani K. 2010. Palynology of the Paravani Tomb Mound (Earle Bronze Age). *Vegetation History and Archaeobotany* 19, 469-478.
- Kvavadze E., Bitadze L., Narimanishvili G., Kakhiani K., Jalabadze M., Koridze I., Rukhadze L., Chichinadze M., Martkoplshvili I. and Laliashvili Sh. 2010. The Bronze Age first-kit, according to palaeobotanical data from archaeological sites in Georgia. In *Abstract of 15th Symposium of IWGE, Wilhelmshaven*. Germany, 54.
- Kvavadze E. and Shatberashvili V. 2010. Results of palynological study of organic remains from Khovle Tombs. *Bulletin, Natural Sciences and Prehistory Section 2*. Tbilisi: Georgian National Museum, 54-74.

- Kvavadze E. and Davadze M. 2014. *The results of palynological researches of the content of the vessel from Pichvnari Necropolis (Grave №339) (= Iberia-Colchis 10)*. Tbilisi: Georgian National Museum Press, 111-117 (in Georgian).
- Leto C., Tuttolomondo T., LaBella S. and Licata M. 2013. Ethnobotanical study in the Madonie Regional Park (Central Sicily, Italy) – Medicinal use of wild shrub and herbaceous plant species. *Journal of Ethnopharmacology* 146, 90-112.
- Li D. I. and Xing F. 2016. Ethnobotanical study on medicinal plants used by local Hoklos people on Hainan Island, China. *Journal of Ethnopharmacology* 194, 358-368.
- Lim T. K. 2012. *Edible Medicinal and Non-Medicinal Plants Vol. 4*. Heidelberg: Springer.
- Lim T. K. 2013. *Edible Medicinal and Non-Medicinal Plants Vol. 5*. Heidelberg: Springer.
- Lim T. K. 2016a. *Edible Medicinal and Non-Medicinal Plants Vol. 10*. Heidelberg: Springer.
- Lim T. K. 2016b. *Edible Medicinal and Non-Medicinal Plants Vol. 12*. Heidelberg: Springer.
- Liu K.C.S., Yang S.L., Roberts M.F., Elford B.C. and Phillipson J.D. 1992. Antimalarial activity of *Artemisia annua* flavonoids from whole plants and cell cultures. *Plant Cell Reports* 11, 637-640.
- Lucas A. and Harris J.R. 1962. *Ancient Egyptian Materials and Industries*. Mineola, NY: Dover.
- Mannan M.M., Maridass M. and Victor B. 2008. A Review on the Potential Uses of Ferns. *Ethnobotanical Leaflets* 12, 281-285.
- Marwat S.K., Hashimi M., Khan K., Khan D.I. and Khan I. 2012. Barley (*Hordeum vulgare* L.) A Prophetic Food Mentioned in Ahadith and its Ethnobotanical Importance. *American-Eurasian Journal of Agriculture & Environmental Science* 12/7, 835-841.
- Maisurashvili N., Dighmelashvili K., Mshvildadze M., Tsotselia M., Koberidze K., Dzhananashvili N. and Chaduneli N. 2018. *Pikrisgora Cemetery*. Tbilisi: National Agency for Cultural Heritage Preservation of Georgia Press.
- Mc Govern P., Jalabadze M., Batiuk S., Callahan M.P., Smith K.E., Hall G. R., Kvavadze E., Maghradze D., Rusishvili N., Bouby L., Failla O., Cola G., Mariani L., Boaretto E., Bacilieri R., This P., Wales N. and Lordkipanidze D. 2017. Early Neolithic wine of Georgia in the South Caucasus. *Proceedings of the National Academy of the United States of America* 114(48), E10309–E10318. doi:10.1073/pnas.1714728114.
- Menale B. and Muoio R. 2014. Use of medicinal plants in the South-Eastern area of the Partenio Region Park (Campania, Southern Italy). *Journal of Ethnopharmacology* 153, 297-307.
- Meuninck J. 2013. *Edible Wild Plants and Useful Herbs*. USA: Morris Book Publishing.
- Mikaili P., Sharifi M., Sarahroodi S. and Shayegh J. 2012. Pharmacological review of medicinal trees spontaneous in Iran: a historical and modern study. *Advances in Environmental Biology* 6, 165-175.
- Moore P. D., Webb J. A. and Collinson M. E. 1991. *Pollen Analysis*. Oxford: Blackwell Scientific Publications.
- Mortensen J. L. 2014. The Implications of Content Analysis for the Interpretation of Unguentaria in Museum Collections. *Theses and Dissertations* 506. <https://dc.uwm.edu/etd/506>
- Mozhui L., Kakati L. N. and Meyer-Rochow V. B. 2021. Entomotherapy: a study of medicinal insects of seven ethnic groups in Nagaland, North-East. *Indian Journal of Ethnobiology and Ethnomedicine* 17, 17. <https://doi.org/10.1186/s13002-021-00444-1>

- Mueller M.S., Runyambo N., Wagner I., Borrmann S., Dietz K. and Heide L. 2004. Randomized controlled trial of a traditional preparation of *Artemisia annua* L. (Annual Wormwood) in the treatment of malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 98/5, 318-321.
- Najafian Y., Hamed S. S., Farshchi M. K. and Feyzabadi Z. 2018. *Plantago major* in Traditional Persian Medicine and modern phytotherapy: a narrative review. *Electron Physician* 10/2, 6390-6399. doi:10.19082/6390
- Papp N., Czégényi D., Hegedűs A., Morschhauser T., Quave C.L., Cianfaglione K. and Pieroni A. 2014. The uses of *Betula pendula* Roth among Hungarian Csángós and Székelys in Transylvania, Romania. *Acta Societatis Botanicorum Poloniae* 83/2, 113-122.
- Pedreiro S., da Ressurreição S., Lopes M., Cruz M.T., Batista T., Figueirinha A. and Ramos F. 2021. *Crepis vesicaria* L. subsp. *Taraxacifolia* Leaves: Nutritional Profile, Phenolic Composition and Biological Properties. *International Journal of Environmental Research and Public Health* 18, 151. <https://doi.org/10.3390/ijerph18010151>
- Piperno DR. 2006. *Phytoliths: A Comprehensive Guide for Archaeologists and Paleoecologists*. New York: Alta Mira Press.
- Poljšak N. and Kočevcar Glavač N. 2021. *Tilia* sp. Seed Oil – Composition, Antioxidant Activity and Potential Use. *Applied Sciences* 11/11, 4932. <https://doi.org/10.3390/app11114932>
- Quattrocchi U. 2012. *CRC World Dictionary of Medicinal and Poisonous Plants*. New York: Taylor & Francis Group, LLC. pp 4017.
- Richter H., Grosser D., Heinz I. and Gasson P. 2004. Iawa list of microscopic features for softwood identification by an Iawa Committee. International Association of Wood Anatomists at the National Herbarium Nederland, Leiden. *The Netherland, Iawa Journal* 25/1, 1-70.
- Souter K. 1995. *Cure Craft: Traditional Folk Remedies and Treatment from Antiquity to the Present Day*. USA: Saffron Walden C. W. Daniel.
- Saginashvili M. 1970. *Glass vessels from Urbnisi Cemetery*. Tbilisi: Metsniereba (in Georgian).
- Swanston-Flatt S. K., Day C., Bailey C. J. and Flatt P. R. 1990. Traditional plant treatments for diabetes. Studies in normal and streptozotocin diabetic mice. *Diabetologia* 33/8, 462-464.
- Taib M., Rezzak Y., Bouyazza L. and Lyoussi B. 2020. Medicinal Uses, Phytochemistry, and Pharmacological Activities of *Quercus* Species. *Evidence-Based Complementary and Alternative Medicine* vol. 2020, Article ID 1920683. <https://doi.org/10.1155/2020/1920683>
- Tetik F., Civelek S. and Cakilcioglu U. 2013. Traditional uses of some medicinal plants in Malatya (Turkey). *Journal of Ethnopharmacology* 146, 331-346.
- Tilford G. L. 1997. *Edible and Medicinal Plants of the West*. USA: Mountain Press Publishing Company.
- Tvalchrelidze Z. and Kvavadze E. 2016. Results of interdisciplinary Research of Artifacts of the “Qvabebi” Monastic – Settling Complex (in Georgian). *Bulletin of the Georgian National Museum* 7, 63-86.
- Trivedi N. and Singh B. 2018. *Chenopodium album*: A miraculous treasure of therapeutic specularities. *Journal of Advanced Scientific Research* 9/1, 14-19.

- van Geel B. 1998. In M. L. van Hove and M. Hendrikse (eds), *A Study of Non- pollen Objects in Pollen Slides (The Types as Described by Dr Bas Van Geel and Colleagues)*. Utrecht: Laboratory of Palynology and Palaeobotany.
- van Geel B., Buurman J., Brinkkemper O., Schelvis J., Aptroot A., van Reenen G. and Hakbijl T. 2003. Environmental reconstruction of a Roman Period settlement site in Uitgeest (The Netherlands), with special reference to coprophilous fungi. *Journal of Archaeological Science* 30, 873-883.
- van Geel B. and Aptroot A. 2006. Fossil ascomycetes in Quaternary deposits. *Nova Hedwigia* 82, 313-329.
- Wiert C. 2006. *Ethnopharmacology of medicinal plants Asia and the Pacific*. Totowa, New Jersey: Humana Press.
- Yamakawa M. 1998. Insect antibacterial proteins; Regulatory mechanisms of their synthesis and a possibility as new antibiotics. *Journal of Sericultural Science of Japan* 67/3, 163-182.
- Zheleva-Dimitrova D., Zhelev Slavov I. and Dimitrova I. 2011. Antioxidant Activity of Some Carduus Species Growing in Bulgaria. *Free Radicals and Antioxidants* 1/4, 15-20.

