Advanced Course on Tissue Remodeling ACTR'04 - (pp.93-110) - Warsaw, 2004.

Clinical Evaluation of Bone Remodeling and Related Disorders

WOJCIECH GLINKOWSKI *,†

* Department of Orthopaedics and Traumatology of Locomotor System¹ "TeleOrto" Center of Excellence Medical University, Lindleya 4 02-005 Warsaw, Poland

[†] Department of Anatomy, Center of Biostructure Research² Medical University, Chałubińskiego 5 02-005 Warsaw, Poland

The bone remodeling defines a mechanism of bone replacement in the skeleton. The skeleton undergoes that process continuously leading to the replacement of the used bone by a newly formed one. The remodeling mechanisms are responsible for accommodation to calcium fluctuations and also for bone response to physical activity. Qualitative or quantitative remodeling disorders have been recognized. The abnormal bone remodeling is observed in Paget's disease leading to pathologic bone formation as well as in osteoporosis, where the process is imbalanced. Clinical features, bone quantity measurements, radiographic images and biochemical markers clearly describe bone status allowing diagnosing and monitoring metabolic bone diseases.

1. Introduction

The mechanism of bone replacement in the skeleton is known as bone remodeling. The skeleton undergoes that process continuously. The remodeling cycle involves the interaction of cells of osteoblastic and osteoclastic lineage and is regulated by both systemic hormones and local factors [1, 2]. Internal secondary bone remodeling of cortex is resulting in the production of

¹⁾Head of the Department: Professor Andrzej Górecki MD, PhD

²⁾Head of the Department: Professor Bogdan Ciszek MD, PhD

Haversian bone, in which most of the bone is occupied by secondary osteones (Haversian systems), or interstitial lamellae.

The reason for replacement is to preserve the functional capacity of bone [3]. Considering the primary mechanical function of bone, the cortical bone carries the mechanical load throughout the diaphyses of long bones. However, load transmission along bone ends is shared to thin cortical bone and well developed peripheral cancellous bone.

The bone is being resorbed and deposited at the same time, often almost continuously. The remodeling process begins "in utero". Burton et al. [4] pointed out that by the 6 month secondary remodeling is going on intensively in the human fetus. It seems extraordinarily unlikely that this remodeling is required by the metabolic needs of the fetus. The purposes of secondary remodeling are to release the needed ions into the circulation, to participate in calcium and phosphate metabolism, hematopoesis support, and mainly growth and bone replacement for used bone units throughout life [5, 6]. The remodeling mechanisms allow the bone to accommodate to calcium fluctuations. This mechanism is responsible also for slow elimination of bone in response to the age-related decline in physical activity. Considering aging, the main purpose of remodeling is to prevent degradation of aging bone.

Another specific function of remodeling is to recover the bone with its normal mechanical function after fracture, defect, osteotomy or other bone pathology. Bone damage as fracture, microfracture, fatigue microdamage forces bone to activate repair mechanisms including remodeling.

The remodeling evolves typically through various phases of activity, followed by a quiescent stage. Phases of remodelling in the case of trabecular bone are schematically depicted in Fig. 1.

Bone replacement begins with osteoclastic resorption followed soon after by osteoblastic formation. However, bone resorption and formation are



FIGURE 1. Simplified bone remodeling cycle

regarded as independent processes, but they seems to be linked as "basic metabolizing units" (BMU) described by Frost (later known as "basic multicellular units") [7,8]. Well developed BMU form the cutting cone or hemicone which consists of a group of osteoclasts in front of the cone, osteoblasts behind forming the closing cone or hemicone, newly forming blood vessels, and connective tissue [9]. The BMU excavates and refills a tunnel inside the cortical bone or across the surface of cancellous bone. The BMU in cortical bone travels circa $20 \,\mu\text{m}/\text{day}$ for about $4000 \,\mu\text{m}$. In cancellous bone BMU's speed is about one half than in cortical bone. Frost [7,8] originally described "activation" as the arising of a new BMU. Its frequency was defined as the birthrate of new BMUs in a unit amount of bone [10,11]. The activation stimulates precursor cells to begin dividing to produce the new cells that comprise the new BMU. The activated cells belong to the population of mesenchymal cells.

In addition to the systemic calcium-regulating hormones, parathyroid hormone, 1,25-dihydroxy vitamin D and calcitonin, sex hormones play an important role for bone remodeling. Estrogen has been identified as the major inhibitor of bone resorption in both men and women. Androgen is an important factor not only as an estrogen source, through the aromatase activity, but also for its direct stimulating bone formation effect. The effects of sex hormones may be mediated by their alteration of local cytokines, prostaglandins and growth factors secretion. The action of sex hormones is also modulated by the level of sex hormone-binding globulin in the circulation. A more accurate evaluation of these effects has been made possible by the development of new methods of measuring bone mineral density and biochemical markers related to rates of bone formation and resorption, as well their influence on risk of osteoporotic fractures [12–33].

2. Qualitative Pathology of Bone Remodeling: Paget's Disease

A remarkable qualitative disorder of bone remodeling is observed in Paget disease [34, 35]. The disease holds the name after an English surgeon, Sir James Paget who described the clinical course of the disease he named *osteitis deformans*. The disease is a relatively common disorder in middle-aged and elderly patients, characterized by excessive and abnormal remodeling of bone. During Paget's disease enhanced resorption of bone is observed. It is performed by giant multinucleated osteoclasts with formation of disorganized

woven bone by osteoblasts. The excessive remodeling gives rise to bones that are extensively vascularized, weak, enlarged, and deformed with subsequent complications. In this disorder, the osteoclasts become abnormally activated. The viral infection is considered as possible cause of the disorder [36], where the bone produces a bizarre and irregular pattern of resorption, to which there is usually an intense osteoblastic response with irregular new bone formation often in the form of woven bone.

Thus, in Paget disease bone density may be increased. Because of the irregular architecture, the Pagetic bone strength decreases and pathologic fractures may occur. There is a genetic component considered in Paget disease etiology [35, 37, 38]. It may be linked to an osteosarcoma tumor suppressor gene [39]. This could account for the increased risk of osteosarcoma in patients with Paget disease.

The remodeling cycle begins with raise of excessive osteoclastic activity with resorption of normal bone by giant multinucleated cells. Osteoblasts respond intensively producing increased amount of disorganized bone with vascular, primitively woven bone and connective tissue reaction. The repeating osteoclastic and osteoblastic activity with bone destruction and formation causes a high degree of bone turnover, and finally abnormal bone production. Osteoclastic activity may decrease and osteoblastic activity also declines after a variable time. Normal-appearing lamellar bone may partially replace immature woven bone. Paget disease typically consists of the following three phases: osteolytic, mixed (osteolytic and osteoblastic) and finally sclerotic. The sequence of stages is variable. Each skeletal lesion is characterized by its own rate of progression. At any time instant, multiple stages of the disease may be demonstrated in different skeletal regions. Clinical features of Paget's disease consist of bony deformities, such as an enlarged skull, spinal kyphosis, and bowing of the long bones of the extremities. Bone angulations and deformity may affect joints with resulting pain and decreased range of motion (ROM). On plain radiographs typical expanding lytic lesions, transverse lucent areas or osteoporosis circumscripta, thickened cortices, sclerotic changes, and bone expansion with coarse disorganized trabecular patterns are seen (Fig. 2).

Radiographic features are diagnostic with an initial osteolytic phase, commonly in the skull and tubular bones, followed by an osteosclerotic phase that is most frequent in the axial skeleton and pelvis. Pagetic long bones looks enlarged with increased radiodensity and trabeculations. Paget disease



FIGURE 2. Radiograms of (a) normal long bone and (b) pagetic bone (analogous region)

typically affects the vertebral bodies and posterior elements. The picture of Paget disease shows enlarged coarse trabeculae combined with the prominent radiodense peripheral contour of the vertebral body and homogenous increase in osseous density in the vertebral body (ivory vertebra). Computer tomography (CT) scanning and magnetic resonance (MR) imaging may be useful for preoperative planning in selected cases. Bone scanning is the most sensitive test for evaluating the extent of lesions in Paget disease. Additional Paget disease specific scintigraphic spinal signs described [40] as the clover or heart signs may aid diagnosis, serve for morbidity prevention, and costs reduction. Histologic findings of Paget disease are marked by disordered areas of resorption and increased number of overly large osteoclasts in the initial osteolytic phase. These abnormal osteoclasts may contain many nuclei. New bone matrix and formation of woven bone is observed during subsequent osteoblastic phase. The histologic hallmark of Paget's disease is seen as joined in a jigsaw or mosaic pattern of many small irregularly shaped bone fragments. These bone fragments are produced by repeated episodes of bone removal and formation. When the osteoblastic phase predominates during

TABLE 1. Biochemistry of bone disorders

	ALP	Ca	Phos.	PTH
Hyperparathyroidemia	Î	Ť	\downarrow	Ť
Osteomalacia	Î	(\downarrow)	Ļ	(\uparrow)
Osteoporosis	N	Ν	Ν	Ν
Paget's	Ţ	N	N	Ν
Bone Metastases	1	(\uparrow)	Ν	(\downarrow)

the disease progresses, more compact and dense bone appears by excessive abnormal bone formation. The Pagetic bone is highly vascularized, rough and fibrous with loose connective tissue filling the marrow spaces. The hypervascularity consists of an increased number of patent capillaries and dilated arterioles, as well as of larger venous sinuses. The normal trabecular appearance is distorted in Pagetic bone with a mosaic pattern of irregular cement lines joining areas of lamellar bone. Pagetic bone shows no tendency to form Haversian systems or to center on blood vessels. The osteoblastic activity diminishes when an osteosclerotic or burned-out phase predominates. The new bone is disordered, poorly mineralized, and lacks structural integrity. Biochemical findings [41] may reveal elevated alkaline phosphatase levels of bone origin, due to increased osteoblastic activity and bone formation.

Procollagen I N-terminal peptide (PINP) recently has emerged as a sensitive serum marker for bone formation. Osteocalcin measurements are usually within the reference range. Levels of urinary hydroxyproline (a product of collagen breakdown), which reflect increased osteoclastic activity and bone resorption, are elevated.

Approximately 20–30% of total hydroxyproline levels are from bone resorption. Measurement of the urinary excretion of bone-specific pyridinium collagen cross-links (urinary pyridinoline collagen cross-link assay) has been found to be a sensitive and specific index of bone resorption. Additionally, levels of excreted bone-specific pyridinium collagen cross-links may be better indicators of bone resorption and response to treatment than the hydroxyproline assay. Urinary N-telopeptide (NTX) and alpha-C telopeptide (CTX) have emerged recently as sensitive biochemical markers for bone resorption. In active Paget disease an abnormally high alpha-CTX: beta-CTX ratio is present. Serum total acid phosphatase (an osteoclastic enzyme) may be elevated in active Paget disease. Increased level of serum total acid phosphatase is observed also in the presence of metastatic prostate carcinoma. Serum calcium and phosphate levels and urinary excretion of calcium should remain within the reference range in patients with Paget disease.

3. Quantitative Alteration of Bone Remodeling: Osteoporosis

Primary osteoporosis is by far the most common metabolic disorder of the skeleton [42], namely expressed as quantitative pathology of bone remodeling. The disease is defined as a skeletal disorder characterized by compromised bone strength that increases the risk of fracture. Osteoporosis has been di-

vided into type 1, or postmenopausal osteoporosis, and type 2, or senile osteoporosis, on the basis of possible differences in etiology. Well known as an illness of older women, osteoporosis may also affect men [43, 44]. Studies of Riggs et all. have suggested that estrogen deficiency is important for the pathogenesis of both types of osteoporosis and in both men and women [45].

The bone strength primarily reflects the integration of bone density and quality [46]. Bone density is expressed as grams of mineral per area or volume, and bone quality is defined as the architecture, turnover, damage accumulation and mineralization. There is at present no accurate measure of bone strength. Bone mineral density (BMD) is a surrogate measure accounting for about 70% of the bone strength. The World Health Organization (WHO) has defined osteoporosis as a bone density 2.5 SDs (standard deviations) below the mean for young adult women. Osteoporosis results from an imbalance of bone remodeling, in which bone resorption outstrips bone formation. The net loss of bone matrix renders bones weaker and more susceptible to fracture, with the fracture risk doubling for every 10 percent bone loss.

Fuller Albright more than 60 years ago [47] pointed out the importance of estrogen in maintaining calcium homeostasis in the postmenopausal woman. Since that time more data demonstrates that hormone replacement (estrogen with or without progesterone) reduces bone turnover and increases bone mass [48]. Recent studies provide stronger evidence of the association between low estradiol concentrations and low bone mass. Ettinger et all. have demonstrated that the lowest estradiol levels in postmenopausal women (i.e. $< 5 \,\mathrm{pg/ml}$) are associated with the lowest bone mineral density and the greatest likelihood of fracture [49]. Low level of estrogen, in some animal models, was associated with rise of IL-6 interleukin synthesis by stromal and osteoblastic cells. Thus, the estrogen may regulate the transcriptional activity of the IL-6 promoter [50]. In other studies changes in tumor necrosis factor (TNF), IL-11 and IL-1 were found as associated with increased bone resorption [51]. Recently receptor activator of NF- κ B ligand (RANKL), identified as a major regulator of osteoclast recruitment, may be necessary for full activation of remodeling.

Manolagas et al. suggest that enhanced bone resorption may lead to bone loss from estrogen deprivation caused by negative ratio of bone formationbone resorption rate [52]. The physically inactive older woman without estrogen replacement therapy is at extremely high risk of bone loss and subsequence problem fractures.

http://rcin.org.pl

BIDLIOTE

55765



FIGURE 3. Radiograms of (a) normal hip bones (early osteoarthritis), (b) pagetic hip bones, (c) trochanteric fracture and (d) femoral neck fracture; Note thickness of cortical bone of the femoral diaphysis. Adequate cortical thickness is seen on picture (a) and thin but sufficient on picture (b), very thin on picture (c) and moderate on (d).

Dietary calcium deficiency, leading to secondary hyperparathyroidism, plays an important role in the pathogenesis osteoporosis. The average calcium intake of elderly women is between 700 and 800 mg/day [53, 54]. Secondary hyperparathyroidism is assured if vitamin D intake is also suboptimal (serum levels of 25 OH vitamin D < 25 ng/ml). PTH stimulates osteoblasts and provokes the remodeling sequence including the elaboration of several cytokines that accelerate bone resorption. Overall this leads to further uncoupling in the bone remodeling cycle, and significant bone loss. Declining serum levels of vitamin D stimulate PTH release and increases bone turnover. LeBoff et al. reported that over 50% of elders with a hip fracture were vitamin D deficient [55]. Chronic elevations in PTH secretion due to primary or tertiary hyperparathyroidism have been associated with low bone mass at several skeletal sites including the radius.

Bone strength is affected by its mass, microarchitecture, macrogeometry, and rate of turnover. Measurements of BMD made at the hip predict hip fracture better than measurements made at other sites [56, 57, 58]. BMD measurement at the spine predicts spine fracture better than measurements at other sites.

Newer measures of bone strength, such as ultrasound, have been established. Studies using quantitative ultrasound (QUS) of the heel have predicted hip fracture and all nonvertebral fractures nearly as well as dual-energy X-ray absorptiometry (DXA) at the femoral neck. Quantitative ultrasound and DXA at the femoral neck offer independent information about fracture risk. Both of these tests predict hip fracture risk better than DXA at the lum-

bar spine. Bone strength is also affected by the rate of remodeling that can be assessed with use of markers of bone turnover in the blood or urine [59].

The clinical use of biochemical markers of bone turnover for the diagnostics, management and monitoring of osteoporosis has been investigated for several years [60, 61]. It is possible to predict fracture risk in an untreated patient population by measure of bone turnover markers [62–65]. Some clinical investigations have shown that biochemical markers of bone turnover are able to predict the rise of BMD and the response of an individual to therapy (hormone replacement therapy—HRT, antiresorptive drugs—raloxifene or alendronate) [66–75]. The decrease of bone markers was associated with increased fracture incidence in patients treated with raloxifene, [72] risedronate, [28] and alendronate [29]. Biochemical markers of bone turnover may be useful in monitoring the progression of disease in an individual. The response to pharmacologic therapy is observed earlier and more pronounced than changes in BMD [30]. Additionally, patient monitoring at early treatment period has the potential to encourage continued treatment compliance and identify individuals who are not responding to treatment.

Markers of bone formation include bone-specific alkaline phosphatase, osteocalcin (bone Gla-protein), procollagen I carboxy (PICP), and N-terminal (PINP) extension peptides. Markers of bone resorption include urinary levels of pyridinolines (Pyr or Pyralink), deoxypyridinolines (D-Pyr or Pyrilink-D), serum and urine levels of type I collagen telopeptides (C-telopeptide products (CTX), and N-telopeptide to helix (NTX). Before the changes in bone mineral density can be detected, the level of bone turnover markers may identify changes in bone remodeling within a relatively short time interval (several days to months) [41].

Bone remodeling at menopause is accelerated with increase of bone formation but the rate is inadequate to replace the bone lost by resorption. Observed imbalance may represent a defect in osteoblast function or loss of template from excessive resorption with perforation of trabecular plates and removal of endosteal cortical bone. The defect in osteoblast function could be the consequence of cellular ageing. The impaired osteoblast function may also be the result of a decrease in the synthesis or activity of systemic and local growth factors.

In pathogenesis of osteoporosis one of the most difficult challenges remaining in the field is the determination of the influence of local factors. Identification of specific factors may lead to exciting new approaches to di-

agnosis and therapy of osteoporosis [76]. The monitoring of osteoporotic patient, require the measurement of biochemical markers of bone turnover, as an accessible tool for daily clinical practice. The measurements achieved need to be interpreted in the context of a complete osteoporotic patient evaluation. An assessment of bone turnover status, bone mineral density and individual osteoporotic fracture risk factors provide information useful for monitoring the response to osteoporosis therapy, requiring further investigations.

4. Conclusions

Qualitative and quantitative disturbances may affect the process of bone remodeling, which is responsible for the bone replacement in the skeleton. Two diseases were selected as examples of bone remodeling disturbances. Qualitative impairment of bone remodeling observed in Paget's disease is considered as abnormal bone producing disorder. The osteoporosis with its imbalanced bone remodeling has an unequal resorption to formation ratio what leads to decrease of bone quantity. Clinical features, bone densitometry, and biochemical markers clearly describe bone status including remodeling. Those tests allow diagnosing and monitoring of metabolic bone diseases. They are the most important clinical factors predicting fracture risk.

References

- 1. A.M. KENNY and L.G. RAISZ, Mechanisms of bone remodeling: implications for clinical practice, J. Reprod. Med. 47(suppl. 1):63-70, 2002.
- L.G. RAISZ, Physiology and pathophysiology of bone remodeling, Clin. Chem., 45(8 Part 2): 1353-1358, 1999.
- J.D. CURREY, The many adaptations of bone, J. Biomech., 36:1487– 1495, 2003.
- P. BURTON, C. NYSSEN-BEHETS, and A. DHEM, Haversian bone remodeling in the human fetus, Acta Anatomica, 135:171–175, 1989.
- A.M. PARFITT, Problems in the application of in vitro systems to the study of human bone remodeling, Proc. Workshop on Human Models of Skeletal Aging, NIH, February 1994, Calcif. Tissue Int., 56(suppl. 1): S5– S7, 1995.

- A.M. PARFITT, Skeletal heterogeneity and the purposes of bone remodeling: implications for the understanding of osteoporosis, [in:] R. Marcus, D. Feldman, and J. Kelsey, [eds.] Osteoporosis, 2nd Ed., Academic, San Diego, CA 2001.
- H.M. FROST, Skeletal structural adaptations to mechanical usage (SATMU): 2. Redefining Wolff's law: the remodeling problem, Anat Rec., 226(4):414-422, 1990.
- H.M. FROST, Bone "mass" and the "mechanostat": a proposal, Anat. Rec., 219(1):1-9, 1987.
- A.M. PARFITT, Osteonal and hemi-osteonal remodeling: The spatial and temporal framework for signal traffic in adult human bone, J. Cell Biochem., 55:273–286, 1994.
- H.M. FROST, Tetracycline-based histological analysis of bone remodeling, Calcif. Tissue Res., 3:211-237, 1969.
- C.J. HERNANDEZ, S.J. HAZELWOOD, and R.B. MARTIN, The relationship between basic multicellular unit activation and origination in cancellous bone, Bone, 25(5):585–587, 1999.
- J.A. KANIS and C.C. GLUER, An update on the diagnosis and assessment of osteoporosis with densitometry, Committee of Scientific Advisors, International Osteoporosis Foundation, Osteoporos. Int., 11:192–202, 2000.
- L.J. MELTON III, E.J. ATKINSON, W.M. O'FALLON, H.W. WAHNER, and B.L. RIGGS, Long-term fracture prediction by bone mineral assessed at different skeletal sites, J. Bone Miner. Res., 8:1227–1233, 1993.
- P. RAVAUD, J.L. RENY, B. GIRAUDEAU, R. PORCHER, M. DOUGA-DOS, and C. ROUX, *Individual smallest detectable difference in bone min*eral density measurements, J. Bone Miner. Res., 14:1449–1456, 1999.
- D.M. BLACK, S.R. CUMMINGS, H.K. GENANT, M.C. NEVITT, L. PALERMO, and W. BROWNER, Axial and appendicular bone density predict fractures in older women, J. Bone Miner. Res., 7:633–638, 1992.

- S.L. HUI, C.W. SLEMENDA, and C.C. JOHNSTON JR., Baseline measurement of bone mass predicts fracture in white women, Ann. Intern. Med., 111: 355-361, 1989.
- 17. P.D. DELMAS, How does antiresorptive therapy decrease the risk of fracture in women with osteoporosis?, Bone, 27:1–3, 2000.
- S.R. CUMMINGS, D.B. KARPF, F. HARRIS, H.K. GENANT, K. EN-SRUD, A.Z. LACROIX, and D.M. BLACK, Improvement in spine bone density and reduction in risk of vertebral fractures during treatment with antiresorptive drugs, Am. J. Med., 112:281–289, 2002.
- H.K. GENANT, K. ENGELKE, T. FUERST, C.C. GLUER, S. GRAMPP, S.T. HARRIS, M. JERGAS, T. LANG, Y. LU, S. MAJUMDAR, A. MATHUR, and M. TAKADA, Noninvasive assessment of bone mineral and structure: State of the art, J. Bone Miner. Res., 11:707-730, 1996.
- P.D. DELMAS, R. EASTELL, P. GARNERO, M.J. SEIBEL, and J. STEPAN The use of biochemical markers of bone turnover in osteoporosis, Committee of Scientific Advisors of the International Osteoporosis Foundation, Osteoporos. Int., 11(Suppl. 6): S2-S17, 2000.
- A.C. LOOKER, D.C. BAUER, C.H. CHESNUT III, C.M. GUNDBERG, M.C. HOCHBERG, G. KLEE, M. KLEEREKOPER, N.B. WATTS, and N.H. BELL, *Clinical use of biochemical markers of bone remodeling: Cur*rent status and future directions, Osteoporos. Int., 11: 467–480, 2000.
- 22. P. GARNERO, E. HAUSHERR, M.-C. CHAPUY, C. MARCELLI, H. GRANDJEAN, C. MULLER, C. CORNIER, G. BREART, P.J. MEUNIER, and P.D. DELMAS, Markers of bone resorption predict hip fracture in elderly women: The EPIDOS prospective study, J. Bone Miner. Res., 11:1531-1538, 1996.
- P. GARNERO, E. SORNAY-RENDU, B. CLAUSTRAT, and P.D. DELMAS, Biochemical markers of bone turnover, endogenous hormones and the risk of fractures in postmenopausal women: The OFELY study, J. Bone Miner. Res., 15:1526-1536, 2000.
- 24. N.H. BJARNASON and C. CHRISTIANSEN, Early response in biochemical markers predicts long-term response in bone mass during hormone

replacement therapy in early postmenopausal women, Bone, **26**: 561–569, 2000.

- 25. P.D. DELMAS, P. HARDY, P. GARNERO, and M. DAIN, Monitoring individual response to hormone replacement therapy with bone markers, Bone, 26:553-560, 2000.
- P. GARNERO, P.D. DELMAS, Variability and response of urinary resorption markers to hormone replacement therapy, J. Bone Miner. Res., 14:470-472, 1999.
- B.L. RIGGS, Are biochemical markers for bone turnover clinically useful for monitoring therapy in individual osteoporotic patients?, Bone, 26:551-552, 2000.
- S. LORENC, W. GLINKOWSKI, P. RYDZYŃSKI, and R. LORENC, Effect of height and body weight on the values of ultrasound parameters in healthy female subjects in population of Poland, [in Polish], Pol. Merk. Lek., 28:80-82, 1998.
- R. EASTELL, I. BARTON, R.A. HANNON, A. CHINES, P. GARNERO, and P.D. DELMAS, *Relationship of early changes in bone resorption* to the reduction in fracture risk with risedronate, J. Bone Miner. Res., 18:1051-1056, 2003.
- 30. D.C. BAUER, D.M. BLACK, P. GARNERO, M. HOCHBERG, S.M. OTT, D.L. SCHNEIDER, D. THOMPSON, J. ORLOFF, S. EWING, and P.D. DELMAS, Reduction in bone turnover predicts hip, non-spine, and vertebral fracture in alendronate treated women: The Fracture Intervention Trial, Osteoporos. Int., 13(suppl. 1): S521, 2002.
- L.J. MELTON, S. KHOSLA, E.J. ATKINSON, W.M. O'FALLON, and B.L. RIGGS, *Relationship of bone turnover to bone density and fractures*, J. Bone Miner. Res., **12**:1083–1091, 1997.
- 32. O. JOHNELL, W.H. SCHEELE, Y. LU, J.-Y. REGINSTER, A.G. NEED, and E. SEEMAN, Additive effects of raloxifene and alendronate on bone density and biochemical markers of bone remodelling in postmenopausal women with osteoporosis, J. Clin. Endocrinol. Metab., 87: 985–992, 2002.

- 33. S. SARKAR, J.-Y. REGINSTER, G. CRANS, A. DIEZ-PEREZ, K. PINETTE, and P. DELMAS, *Relationship between changes in biochemical markers of bone turnover and BMD to predict vertebral fracture risk*, J. Bone Mineral Res., **19**(3):394, 2004.
- B.G. MILLS, A. FRAUSTO, Cytokines expressed in multinucleated cells: Paget's disease and giant cell tumors versus normal bone, Calcif. Tissue Int., 61: 16-21, 1997.
- E.S. SIRIS, Paget's disease of bone, J. Bone Miner. Res., 13:1061–1065, 1998.
- A.P. MEE, J.A. DIXON, J.A. HOYLAND, M. DAVIES, P.L. SELBY, and E.B. MAWER, Detection of canine distemper virus in 100% of Paget's disease samples by in situ reverse transcriptase-polymerase chain reaction, Bone, 23:171–175, 1998.
- J.A. KANIS, [ed.], Pathophysiology and Treatment of Paget's Disease of Bone, Second ed., Martin Dunitz Ltd, London 1998.
- E.S. SIRIS, Seeking the elusive etiology of Paget disease: a progress report, J. Bone Miner. Res., 11(11):1599–1601, 1996.
- M.J. NELLISSERY, S.S. PADALECKI, Z. BRKANAC, F.R. SINGER, G.D. ROODMAN, K.K. UNNI ET AL., Evidence for a novel osteosarcoma tumor-suppressor gene in the chromosome 18 region genetically linked with Paget disease of bone, Am. J. Hum. Genet., 63:817-824, 1998.
- 40. D. ROTE'S-SALA, J. MONFORT, A. SOLANO, E. MIRALLES, J. VILA, and J. CARBONELL, The clover and heart signs in vertebral scintigraphic images are highly specific of Paget's disease of bone, Bone, 34:605-608, 2004.
- S. LELLO, A.M. PAOLETTI, S. MIGLIACCIO, G.B. MELIS, Bone markers: biochemical and clinical significance, Aging Clin. Exp. Res., 16(Suppl 3): 33-6, 2004.
- L.G. RAISZ, The osteoporosis revolution, Ann. Intern. Med., 126:458– 462, 1997.
- 43. R. MARCUS, An expanded overview of postmenopausal osteoporosis,
 J. Musculoskelet. Neuronal Interact., 2(3): 195–197, 2002.

- E. SEEMAN, Pathogenesis of bone fragility in women and men, Lancet, 359:1841–1850, 2002.
- 45. B.L. RIGGS, S. KHOSLA, and L.J. MELTON, A unitary model for involutional osteoporosis: estrogen deficiency causes both type I and type II osteoporosis in postmenopausal women and contributes to bone loss in aging men, J. Bone Miner. Res., 13: 763–773, 1998.
- NIH Consensus Development Panel on Osteoporosis Prevention, Diagnosis, and Therapy, March 7-29, 2000: Highlights of the Conference, South. Med. J., 94(6): 569–573, 2001.
- F. ALBRIGHT, Postmenopausal osteoporosis, JAMA, 116:2465-2474, 1941.
- 48. C. CHESTNUST, M. NOTELOVITZ, G. CLARK, B. DRINKWATER, C. ROSEN, N. BELL, S. ENGLISH, C.C. JOHNSTON, D. CAIN, K. FLESS-LAND, and N. MALLINAK, Use of the N-telopeptide of type I collagen to monitor the effect of therapy and predict changes in bone mineral density in postmenopausal women treated with hormone replacement therapy, Am. J. Med., 102:29–37, 1997.
- B. ETTINGER, A. PRESSMAN, P. SKLARIN, D. BAUER, J.A. CUALEY, and S.R. CUMMINGS, Associations between low levels of serum estradiol, bone density and fractures among elderly women: SOF, J. Clin. Endocrinol. Metab., 83: 2239-2243, 1998.
- R.G. JILKA, G.H. GIRSOLE, G. PASSERI, D. WILLIAMS, J. ABRAMS, B. BOYCE, H. BROXMEYER, and S. MANOLOAGAS, *Increased osteoclast* development after estrogen loss: mediation by IL-6, Science, 257:88–91, 1992.
- 51. R. PACIFICI, C. BROWN, and E. PUSCHECK, The effect of surgical menopause and estrogen replacement on cytokine release from human blood monocytes, Proc. Natl. Acad. Sci. USA, 88:5134-5138, 1991.
- S.C. MANOLAGAS and R.L. JILKA, Emerging insights into the pathophysiology of osteoporosis, N. Engl. J. Med., 332: 305–311, 1995.
- Food and Nutrition Board Institute of Medicine, Dietary Reference Intakes for Calcium, Phosphorus, Magnesium Vitamin D and Fluoride, National Academy Press, Washington DC 1997.

- L.D. MCCABE, B.R. MARTIN, G.P. MCCABE, C.C. JOHNSTON, C.M. WEAVER, and M. PEACOCK, *Dairy intakes affect bone density in the elderly*, Am. J. Clin. Nutr., 80(4): 1066-1074, 2004.
- 55. M.S. LEBOFF, L. KOHLMEIER, S. HURWITZ, J. FRANKLIN, J. WRIGHT, and J. GLOWACKI, Occult vitamin D deficiency in postmenopausal US women with acute hip fracture, JAMA, 282:1505–1511, 1999.
- 56. G. GANDOLINI and P.M. SALVIONI, Is BMD measurement an adequate surrogate for anti-fracture efficacy?, Aging Clin. Exp. Res., 16(Suppl. 3): 29–32, 2004.
- 57. M.L. FROST, I. FOGELMAN, G.M. BLAKE, P.K. MARSDEN, and G. COOK JR., Dissociation between global markers of bone formation and direct measurement of spinal bone formation in osteoporosis, J. Bone Miner. Res., 19(11):1797–1804, 2004.
- D. AGNUSDEI, Bone quality, Aging Clin. Exp. Res., 16(Suppl 3):1-2, 2004.
- NIH Consensus Development Panel on Osteoporosis Prevention, Diagnosis, and Therapy, March 7–29, 2000: Highlights of the Conference, South. Med. J., 94(6): 569–573, 2001.
- 60. P.D. DELMAS, R. EASTELL, P. GARNERO, M.J. SEIBEL, and J. STEPAN, *The use of biochemical markers of bone turnover in osteoporosis*, Committee of Scientific Advisors of the International Osteoporosis Foundation, Osteoporos. Int., **11**(Suppl 6): S2–S17, 2000.
- 61. A.C. LOOKER, D.C. BAUER, C.H. CHESNUT III, C.M. GUNDBERG, M.C. HOCHBERG, G. KLEE, M. KLEEREKOPER, N.B. WATTS, and N.H. BELL, *Clinical use of biochemical markers of bone remodeling: Cur*rent status and future directions, Osteoporos. Int., 11: 467–480, 2000.
- 62. P. GARNERO, E. HAUSHERR, M.-C. CHAPUY, C. MARCELLI, H. GRANDJEAN, C. MULLER, C. CORNIER, G. BREART, P.J. MEUNIER, and P.D. DELMAS, Markers of bone resorption predict hip fracture in elderly women: The EPIDOS prospective study, J. Bone Miner. Res., 11:1531–1538, 1996.

- 63. P.L. VAN DAELE, M.J. SEIBEL, H. BURGER, A. HOFMAN, D.E. GROBBEE, J.P. VAN LEEUWEN, J.C. BIRKENHAGER, and H.A. POLS, Case-control analysis of bone resorption markers, disability, and hip fracture risk: The Rotterdam study, BMJ, 312:482–483, 1996.
- P.D. Ross, B.C. KRESS, R.E. PARSON, R.D. WASNICH, K.A. AR-MOUR, and I.A. MIZRAHI, Serum bone alkaline phosphatase and calcaneus bone density predict fractures: A prospective study, Osteoporos. Int., 11:76–82, 2000.
- 65. P. GARNERO, E. SORNAY-RENDU, B. CLAUSTRAT, and P.D. DELMAS, Biochemical markers of bone turnover, endogenous hormones and the risk of fractures in postmenopausal women: The OFELY study, J. Bone Miner. Res., 15:1526–1536, 2000.
- 66. N.H. BJARNASON and C. CHRISTIANSEN, Early response in biochemical markers predicts long-term response in bone mass during hormone replacement therapy in early postmenopausal women, Bone, 26:561–569, 2000.
- P.D. DELMAS, P. HARDY, P. GARNERO, and M. DAIN, Monitoring individual response to hormone replacement therapy with bone markers, Bone, 26:553-560, 2000.
- N.H. BJARNASON and C. CHRISTIANSEN, Early response in biochemical markers predicts long-term response in bone mass during hormone replacement therapy in early postmenopausal women, Bone, 26:561-569, 2000.
- 69. P.D. DELMAS, P. HARDY, P. GARNERO, and M. DAIN, Monitoring individual response to hormone replacement therapy with bone markers, Bone, 26:553-560, 2000.
- R. HANNON, A. BLUMSOHN, K. NAYLOR, and R. EASTELL, Response of biochemical markers of bone turnover to hormone replacement therapy: Impact of biological variability, J. Bone Miner. Res., 13: 1124–1133, 1998.
- P. GARNERO and P.D. DELMAS, Variability and response of urinary resorption markers to hormone replacement therapy, J. Bone Miner. Res., 14:470-472, 1999.

- 72. M.H. BJARNASON, S. SARKAR, T. DUONG, B. MITLAK, P.D. DEL-MAS, and C. CHRISTIANSEN, Six and twelve month changes in bone turnover are related to reduction in vertebral fracture risk during 3 years of raloxifene treatment in postmenopausal osteoporosis, Osteoporos. Int., 12:922-930, 2001.
- M.A. BRAGA DE CASTRO, R. HANNON, and R. EASTELL, Monitoring alendronate therapy for osteoporosis, J. Bone Miner. Res., 14:602–608, 1999.
- 74. P. RAVN, B. CLEMMESEN, and C. CHRISTIANSEN, Biochemical markers can predict the response in bone mass during alendronate treatment in early postmenopausal women, Alendronate Osteoporosis Prevention Study Group, Bone, 24:237-244, 1999.
- 75. P. GARNERO, C. DARTE, and P.D. DELMAS, A model to monitor the efficacy of alendronate treatment in women with osteoporosis using a biochemical marker of bone turnover, Bone, 24:603-609, 1999.
- K.M. PRESTWOOD, C.C. PILBEAM, and L.G. RAISZ, Treatment of osteoporosis, Annual Rev. Med., 46:249–256, 1995.